



Artificial intelligence in antimicrobial stewardship: a systematic review and meta-analysis of predictive performance and diagnostic accuracy

Flavia Pennisi^{1,2} · Antonio Pinto² · Giovanni Emanuele Ricciardi^{1,2} · Carlo Signorelli² · Vincenza Gianfredi³

Received: 1 November 2024 / Accepted: 19 December 2024

© The Author(s), under exclusive licence to Springer-Verlag GmbH Germany, part of Springer Nature 2025

Abstract

The increasing threat of antimicrobial resistance has prompted a need for more effective antimicrobial stewardship programs (AMS). Artificial intelligence (AI) and machine learning (ML) tools have emerged as potential solutions to enhance decision-making and improve patient outcomes in AMS. This systematic review and meta-analysis aims to evaluate the impact of AI in AMS and to assess its predictive performance and diagnostic accuracy. We conducted a comprehensive literature search across PubMed/MEDLINE, Scopus, EMBASE, and Web of Science to identify studies published up to July 2024. Studies included were observational, cohort, or retrospective, focusing on the application of AI/ML in AMS. The outcomes assessed were the area under the curve (AUC), accuracy, sensitivity, specificity, negative predictive value (NPV), and positive predictive value (PPV). We calculated the mean pooled effect size (ES) and its 95% confidence interval (CI) using a random-effects model. The risk of bias was assessed using the QUADAS-AI tool, and the protocol was registered in PROSPERO. Out of 3,458 retrieved articles, 80 studies met the inclusion criteria. Our meta-analysis demonstrated that ML models exhibited strong predictive performance and diagnostic accuracy, with the following results: AUC [ES: 72.28 (70.42–74.14)], accuracy [ES: 74.97 (73.35–76.58)], sensitivity [ES: 76.89; (71.90–81.89)], specificity [ES: 73.77; (67.87–79.67)], NPV [ES: 79.92 (76.54–83.31)], and PPV [ES: 69.41 (60.19–78.63)] across various AMS settings. AI and ML tools offer promising enhancements due to their strong predictive performance. The integration of AI into AMS could lead to more precise antimicrobial prescribing, reduced antimicrobial resistance, and better resource utilization.

Keywords Artificial intelligence · Antimicrobial stewardship · Machine learning · Systematic review · Meta-analysis

Introduction

Antimicrobial resistance (AMR) is one of the most pressing global health threats, contributing to increased morbidity, mortality, and healthcare costs worldwide, leading to over 1.5 million deaths worldwide in 2019 alone [1], with a broader implication of 4.95 million deaths when associated with resistant bacterial infections [2]. The emergence

of resistant pathogens has significantly complicated clinical decision-making, particularly in the context of antimicrobial stewardship (AMS), where the goal is to use antimicrobials judiciously to reduce resistance development while ensuring effective treatment [3]. However, studies have shown that approximately 30–50% of antimicrobial use in hospitals is inappropriate, contributing to the acceleration of resistance [4]. In the context of AMS, the ability to predict resistance patterns using rapid diagnostics is crucial. However, traditional culture-based methods of identifying resistance patterns rely heavily on culture-based diagnostics, which can take up to 48–72 h, delaying optimal therapy. This uncertainty in clinical decision-making increases reliance on empirical, often broad-spectrum antibiotic use, exacerbating resistance problems and negatively impacting patient outcomes [5].

Artificial intelligence (AI) and machine learning (ML) technologies have emerged as promising tools to support AMS by enabling rapid, data-driven predictions of AMR and

✉ Vincenza Gianfredi
vincenza.gianfredi@unimi.it

¹ PhD National Programme in One Health approaches to infectious diseases and life science research, Department of Public Health, Experimental and Forensic Medicine, University of Pavia, 27100 Pavia, Italy

² Faculty of Medicine, University Vita-Salute San Raffaele, Milan, Italy

³ Department of Biomedical Sciences for Health, University of Milan, Via Pascal 36, 20133 Milan, Italy

guiding antimicrobial therapy decisions. ML models, particularly those utilizing ML algorithms, have shown potential in various domains of healthcare, including diagnostic accuracy, predictive analytics, and clinical decision support [5]. In the context of AMS, AI-driven approaches could offer more timely and accurate predictions of antimicrobial resistance, potentially guiding clinicians towards more targeted and appropriate therapies [6].

Despite these advancements, there remains a need to comprehensively evaluate the clinical utility of AI and ML models in AMS. Previous studies have reported various performance metrics, and some preliminary reviews have consolidated these findings [7]. However, the absence of any previous meta-analysis makes it impossible to statistically evaluate the overall predictive performance and diagnostic accuracy of AI and ML tools in this domain, which is essential for understanding their value in real-world clinical settings.

This systematic review and meta-analysis aims to evaluate the impact of AI and ML on antimicrobial stewardship, with a specific focus on their predictive performance and diagnostic accuracy. By analyzing key outcomes, including area under the curve (AUC), accuracy, sensitivity, specificity, positive predictive value (PPV), and negative predictive value (NPV), this study will provide insights into the potential role of AI in improving clinical outcomes in the fight against AMR. Through the analysis of the current literature, we aim to inform future research and the implementation of AI technologies in AMS programs.

Methods

Protocol and registration

This systematic review adhered to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines. The protocol was registered with the International Prospective Register of Systematic Reviews

(PROSPERO), registration number [PROSPERO ID CRD42024567640].

Literature search strategy

A comprehensive literature search was performed across PubMed/MEDLINE, Scopus, EMBASE, and Web of Science (WoS) in July 2024. This systematic review seeks to answer the following question: “What is the impact and effectiveness of AI in AMS aimed at reducing AMR?”. The search strategy incorporated two key elements: artificial intelligence (and synonyms) and antimicrobial stewardship or antimicrobial resistance (and synonyms). Selected keywords, including MeSH terms and Title/Abstract terms, were combined using the Boolean operators “AND” and “OR”. The search strategy was initially developed for PubMed/MEDLINE and subsequently adapted for Scopus, EMBASE, and WoS. The search strategies for each database are detailed in Table S1. Additionally, forward and backward citation searches were performed. No temporal filter was applied.

Eligibility criteria

Inclusion and exclusion criteria were defined based on the Population, Intervention, Comparison, Outcome, and Study design (PICOS) guidelines. Only studies involving patients (both inpatient or outpatient) requiring antimicrobial prescriptions in settings (population) and using ML models in AMS were included. We included studies that developed and, where applicable, validated ML models aimed at improving antimicrobial stewardship practices. Moreover, our primary outcomes included predictive performance metrics (AUC, c-statistic, accuracy, sensitivity, specificity, PPV, NPV, and F1-score). Secondary outcomes included clinical outcomes and improvement in patient health outcomes. Lastly, only observational studies published in English in international, peer-reviewed scientific journals, were considered eligible. A detailed description of the inclusion criteria, defined according to PICOS, is provided in Table 1.

Table 1 Inclusion criteria declined according to Population, Intervention, Comparison, Outcome, and Study design (PICOS) guidelines

| | Inclusion criteria |
|------------------|---|
| Population (P) | Patients who require antimicrobial prescriptions in both inpatient or outpatient settings |
| Intervention (I) | Artificial intelligence or machine learning tools used in antimicrobial stewardship |
| Comparison (C) | Traditional clinical decision-making processes without the aid of artificial intelligence |
| Outcome (O) | Main outcome: measure of the effectiveness of machine learning algorithms to support antimicrobial stewardship programs: AUC, c-statistic, accuracy, sensitivity, specificity, PPV, NPV and F1-score Secondary outcome: clinical outcomes and improvement in patient health outcomes |
| Study design (S) | Observational, including cohort studies, prospective or retrospective |
| Time filter | No temporal filter |

AUC Area under the curve; PPV positive predictive value; NPV negative predictive value

Study selection

Search results were imported into Mendeley for organization and de-duplication. Titles and abstracts were independently screened, in blind, by two reviewers using Rayyan software (<https://rayyan.qcri.org>). Full-text articles were obtained and assessed, in blind by two reviewers against the eligibility criteria. Discrepancies were resolved through discussion or by consulting a third reviewer. A PRISMA flowchart was used to document the study inclusion process, recording the number of articles retained at each screening stage and reasons for exclusion at the full-text screening stage.

Data extraction

Data extraction was performed in blind using a standardized template in Microsoft Excel (Microsoft Excel® for Microsoft 365 MSO, Redmond, WA, USA, 2019). The spreadsheet was initially tested on three randomly selected articles to enhance consistency and agreement among the authors. Extracted data included: study characteristics (author name, year of publication, country where the study took place, study design), population characteristics (demographic data, sample size, hospital setting, type of infection, clinical settings), ML model details (ML methods used, training data sets, number of features, data source i.e. clinical and/or laboratory data), and outcomes (predictors, performance validation, clinical outcomes). Discrepancies in data extraction were resolved through discussion or by consulting a third reviewer.

Risk of bias assessment

The risk of bias in the included studies was assessed independently in blind by two authors using the QUADAS-AI tool [8]. This tool allows for a detailed and structured assessment of various bias domains, including patient selection, index test, reference standard, and type of validation. Discrepancies in assessments will be resolved through discussion and, if necessary, by consulting a third senior reviewer to reach a consensus. This comprehensive assessment is crucial for understanding the reliability and validity of the study findings.

Data synthesis and meta-analysis

Data synthesis was organized into tables to provide an overview of study characteristics and findings. In the meta-analysis, only performance measures with data from at least 5 distinct studies were included. Quantitative analyses were conducted on performance measurements related to validation, where available. A quantitative synthesis of the data was planned if feasible, using random-effects models

to assess the AUC, accuracy, sensitivity, specificity, PPV, and NPV of ML models. Heterogeneity among studies was assessed using I^2 statistics. Publication bias was assessed using funnel plots and Egger's test if at least ten studies were included in the meta-analysis.

ML models were grouped based on functional similarities, considering their approach to data processing, underlying mechanisms and typical applications. This grouping facilitated a more coherent comparison across studies by ensuring that models with comparable characteristics were analysed together. Specifically, the following groups were identified:

- Decision Trees, including Random Forest (RF), Decision Tree (DT), Classification and Regression Trees (CART), and J48 (C4.5).
- Boosted Models, including Extreme Gradient Boosting (XGB), Gradient Boosted Decision Trees (GBDT), Categorical Boosting (CatBoost), Adaptive Boosting (AdaBoost), Boosted Decision Trees (Boosted DT), Boosted Logistic Regression (Boosted LR), and Gradient Boosting Machine (GBM).
- Neural Networks were divided into two categories: generic networks, such as Artificial Neural Networks (ANN), Neural Networks (NN), Backpropagation Neural Networks (Backpropagation NN), and Neural Networks with SHapley Additive exPlanations (NN with SHAP); and recurrent networks, including Long Short-Term Memory (LSTM), Bidirectional Long Short-Term Memory (Bidirectional LSTM), Recurrent Neural Networks (RNN), and Gated Recurrent Unit (GRU).
- Support Vector Machines (SVMs), including standard Support Vector Machines (SVM), SVM with Radial Basis Function kernel (SVM with RBF kernel), Sequential Minimal Optimization (SMO), Linear Support Vector Machines (Linear SVM), Polynomial Support Vector Machines (Polynomial SVM), and SVM C-Support Vector.

Results

Literature search

A total of 3,458 records were identified through searches in PubMed/MEDLINE ($n=992$), Scopus ($n=1,134$), Embase (705), and Web of Science (627) databases. Additionally, two articles were included based on reference screening. Following the initial removal of duplicates ($n=1,825$), a total of 1,633 records underwent screening based on title and abstract. Subsequently, 1,477 records were eliminated due to non-original content and focus on different topics, resulting in 156 records deemed eligible for inclusion. Three

articles [9–11] lacked full-text availability. Following the full texts assessment, 75 records were excluded, including 80 records [12–90]. The study selection process is visually represented in Fig. 1.

Descriptive characteristics of included studies

The systematic review comprised 80 studies, encompassing a total of over 1.3 million patients. All studies were published after the year 2000. The geographic distribution is notably concentrated (Figure S1), with the highest contribution from the USA ($n=20$ studies) [14, 19, 20, 22, 28, 30, 31, 36, 41, 43–45, 61, 62, 66, 67, 69, 72, 89, 90], Taiwan ($n=9$) [13, 32, 39, 50, 59, 60, 78, 83, 84], Israel [21, 35, 53, 56, 68, 73, 82, 91] and Spain [34, 49, 54, 57, 64, 65, 74, 80] with 8 studies each. Most studies (70%, $n=56$) [12, 15–19, 21–30, 32, 35, 38, 39, 46–50, 53–58, 60–62, 64, 65, 68, 70, 72–84, 86–89, 92] were monocentric, conducted at a single

institution (Table 2). Genitourinary infections were the predominant area of research focus, comprising 38.8% ($n=31$) of total studies [12, 13, 16–20, 22, 23, 25, 33, 35, 41, 44, 53, 56–60, 62, 68–71, 75, 76, 83, 86, 91, 92]. Bloodstream infections accounted for 26.3% ($n=21$) of the studies [13, 16, 17, 24, 31, 43, 46, 49, 50, 59, 60, 62–65, 69, 70, 77, 78, 80, 83], followed by respiratory infections (21.3%, $n=17$) [13, 16, 19, 22, 26, 42, 51, 59, 60, 64, 67, 69–71, 75, 85, 92]. The majority of the studies (82.5%, $n=66$) [12–25, 28–30, 32, 34–37, 40, 42, 43, 45–62, 64–69, 71–74, 77, 78, 80–82, 85–92] report data on inpatient (I) populations, while only 2.5% ($n=2$) [27, 33] have outpatient data (O). A further 7.5% ($n=6$) [41, 44, 63, 75, 76, 83] include both inpatient and outpatient data (I/O), 5% ($n=4$) [26, 31, 38, 84] report data on emergency patients, and 2.5% ($n=2$) [39, 70] do not specify this information (NA).

Of the 80 included studies, only 7 [12, 29, 36, 44, 51, 59, 83] used external validation and one [71] employed both

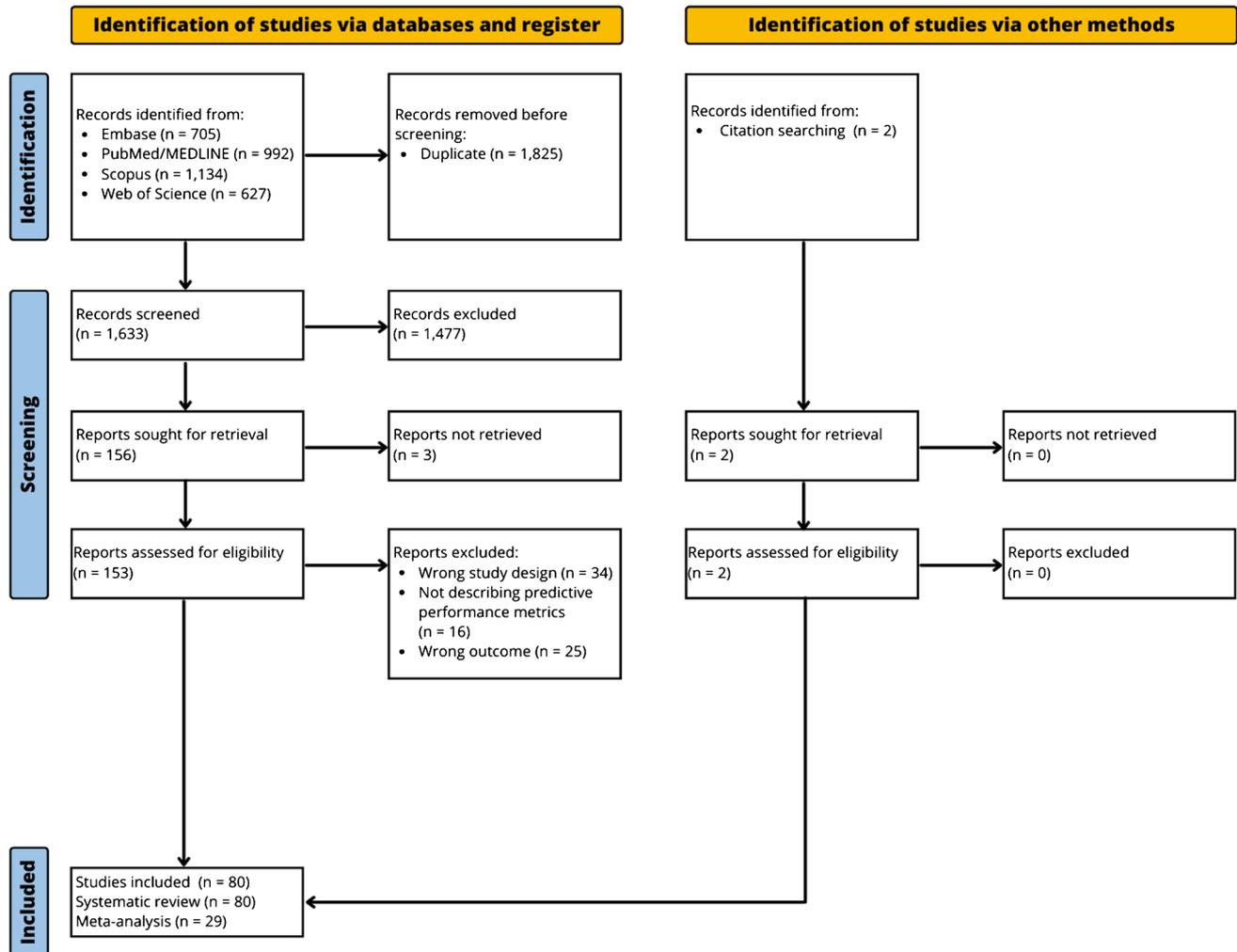


Fig. 1 Flow diagram depicting the selection process

Table 2 Descriptive characteristics of included studies

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|-----------------------|------------|---------|------|-----------------|--|--|--|----|----------------|----------------|--------|----------|
| | | | | | | | | I | UTI | Yes / Internal | No | NA |
| Abu-Aqil G, 2022 [35] | RCS | Israel | 1 | NA | To prove the potential of FTIR spectroscopy combined with ML algorithms for determining the susceptibility of E. coli | Patients with UTI | Test 1798 distinct isolates of E. coli | I | UTI | Yes / Internal | No | NA |
| Abu-Aqil G, 2023 [91] | ERS | Israel | 1 | NA | To assess the effectiveness of IR with XGB ML for rapid E. coli identification and antibiotic susceptibility from urine samples, reducing diagnosis time | Patients with suspected UTIs from whom urine samples were collected to identify E. coli isolates | 1765 E. coli isolate | I | Urinary | Yes / Internal | NA | 16/398 |
| Abu-Aqil G, 2023 [53] | OBS | Israel | 1 | NA | Evaluate using IR with ML for rapid detection of Proteus mirabilis and P. aeruginosa in urine samples and determining their antibiotic susceptibility | Urine samples from patients with UTI | 3446 urine samples analyzed (360 Proteus mirabilis, 353 P. aeruginosa) | I | UTI | NA / NA | No | 469 |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|---------------------------|------------|---------|------|-------------------|---|---|--|-----|------------------------------------|----------------|--------|------------|
| | | | | | | | | I | Urinary | Yes / Internal | No | |
| Abu-Aqil G, 2024 [56] | RCS | Israel | 1 | NA | To assess the feasibility of IR-based ML for rapid, accurate K. pneumoniae identification and antibiotic susceptibility from urine samples | Patients with UTIs, specifically those infected with K. pneumoniae | 2333 patients, 636 with K. pneumoniae infections, and the rest with other UTI-related bacterial infections | | Urinary | Yes / Internal | No | 50–400/469 |
| Agbaria A-H, 2020 [82] | PCS | Israel | 1 | NA | To evaluate the potential of mid-IR combined with ML algorithms for rapid diagnosis of infection etiology (bacterial versus viral) in febrile pediatric oncology patients | Febrile pediatric oncology patients aged 0–18 years treated with chemotherapy | 141 blood samples (50 viral, 71 bacterial, and 20 controls) | I | NA | Yes / Internal | NA | 120–127 |
| Al-khlifeh E-M, 2024 [75] | RCS | Jordan | 1 | Oct 2020-Dec 2022 | To predict the AMR for various antibiotic categories using two ML models and assess the impact of demographic factors on AMR | The dataset included 2893 reports; 72.2% female, age range from less than one week to 102 years | 2893 | I/O | UTI, gastrointestinal, respiratory | Yes / Internal | No | NA |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|-----------------------------|------------|-----------|------|----------------------------|--|--|----------------------------------|----|----------------|----------------|--------|----------|
| Ananda-Rajah M-R, 2017 [85] | CCS | Australia | 3 | NA | Develop an expert system for IMD surveillance using NLP of CT reports, microbiology, and antifungal drug data to improve IMD prediction | Patients from hematology-oncology departments, focusing on high-risk cases for invasive mold disease | 123; 64 with IMD and 59 controls | I | Respiratory | NA / NA | No | NA |
| Ayyildiz H, 2021 [58] | RCS | Turkey | 1 | 2019–2020 | To evaluate if E. coli antibiotic resistance can be predicted using ML models on routine lab data without conventional antibiogram tests | Patients with E. coli infections | 103 | I | UTI | Yes / Internal | No | 12/12 |
| Beaudoin M, 2016 [15] | PCS | Canada | 1 | Feb–Nov 2012; Nov–Dec 2013 | To evaluate the ability of the algorithm to discover rules for identifying inappropriate prescriptions of TZP | Patients monitored by APSS who received at least one prescription of TZP at the Centre Hospitalier Universitaire de Sherbrooke | 421 | I | NA | Yes / Internal | No | NA |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|-----------------------|------------|---------|----------|--------------------|--|---|---|----|--------------------------|---|--------|--------------------------------|
| Bhavani S, 2020 [43] | RCS | USA | 2 | 2007–2018 | To develop and validate ML models to predict blood culture results at the time of order using routine EHR data | Hospitalized adult patients who received a blood culture | Over 250000 blood culture days across both centers | I | Bloodstream | Yes / Internal | Yes | NA |
| Bolton W-J, 2022 [71] | RCS | UK | 1 | June 2017–Jan 2019 | To estimate patients' ICU LOS and mortality outcomes for any given day, under the alternative scenarios of stopping versus continuing antibiotic treatment | Patients with UTI and wound infections from Maccabi Healthcare Services with a positive wound culture record | 18988 patients, associated with 22845 unique ICU stays; train 15991 stays, validation 3427 stays, test 3427 stays | I | UTI | Yes / Internal | No | 43 |
| Bolton W-J, 2024 [62] | OBS | UK | Multiple | 2008–2019 | To develop and evaluate a ML model to predict the appropriateness of switching from IV to oral antibiotic therapy in ICU settings | ICU patients from MIMIC database and eICU Col-laborative Research Data-base who received both IV and oral antibiotic treatments | 8694 unique ICU stays from MIMIC, 1668 from eICU | I | UTI, Respiratory, Sepsis | Yes / Internal (MIMIC), External (eICU) | Yes | Short model: 5, long model: 37 |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|---------------------------|------------|---------|------|-------------------|--|--|---|----|---|----------------|--------|----------|
| Brokowski T-J, 2022 [62] | RCS | USA | 1 | 2001–2012 | To develop and validate ML models predicting antibiotic susceptibility in <i>S. pneumoniae</i> , infections, improving stewardship and optimizing antibiotic selection | Adult patients suspected of having <i>S. pneumoniae</i> infections | 4734 unique ICU admissions, with 7655 antibiotic prescriptions | I | Bloodstream, urine, cerebral spinal fluid, pleural cavity, joints | Yes / Internal | No | 87/4754 |
| Brown D-G, 2023 [45] | PCS | USA | 5 | 2017–2020 | To derive and validate a clinical prediction rule for identifying USA international travelers at risk of acquiring ESBL-PE during travel | USA international travelers ESBL-negative pre-travel who provided stool samples before and after travel | 528 travelers | I | Gastrointestinal | Yes / Internal | Yes | 27 |
| Bystritsky R-J, 2020 [17] | RCS | EUA | 1 | Dec 2015–Aug 2017 | Predict whether antibiotic therapy requires intervention on a given day based on the note from the antimicrobial stewardship team in the patient's record | Hospitalized patients who received at least one antimicrobial tracked by the ASP at University of California, San Francisco MC | 9651; split randomly into derivation (80%) and validation (20%) data sets | I | Bloodstream, UTI | Yes / Internal | Yes | 56 |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|-----------------------|------------|---------|------|-------------------|--|---|---|----|-------------------------------|----------------|----------------|----------|
| | | | | | | | | | | Yes / Internal | Yes / Internal | No |
| Çağlayan Ç, 2022 [28] | RCS | USA | 1 | 2017–2018 | To build a predictive framework for accurate MDRO colonization detection, ensuring high sensitivity and specificity to optimize resource use | Patients admitted to a surgical or medical ICU | 3958 patients; 17.59% MDRO, 13.03% VRE, 1.45% CRE, 7.47% MRSA | I | NA | Yes / Internal | No | 26 |
| Cai T, 2023 [33] | RCS | Italy | 2 | Jan 2012–Dec 2020 | Predicting the clinical efficacy of the empiric antimicrobial treatment in women with recurrent UTI | Patients who had undergone antimicrobial treatment for uncomplicated cystitis | 1043 (725 train, 318 testing) | O | Urinary | Yes / Internal | Yes | NA |
| Candela A, 2022 [80] | OBS | Spain | 1 | 2017–2019 | To develop a MALDI-TOF-based classifying algorithm for discriminating between VRE and VSE faecium isolates | Clinical isolates of Enterococcus faecium from hospitalized patients | 178 clinical isolates (92 vancomycin-susceptible, 31 VanA-resistant, and 55 VanB-resistant) | I | Bloodstream, gastrointestinal | Yes / Internal | No | 18 |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|------------------------|------------|---------|------|-------------------|--|--|---|----|----------------|----------------|--------|----------|
| | | | | | | | | | | Yes / Internal | No | |
| Cavallaro M, 2023 [55] | RCS | UK | 1 | Jan 2010-Oct 2016 | To develop an explainable AI model predicting antimicrobial resistance in hospitals, reducing treatment mismatches, and improving antibiotic stewardship | Inpatients who had GNB isolates from blood and urine cultures, specifically focusing on <i>E. coli</i> , <i>K. pneumoniae</i> , and <i>P. aeruginosa</i> | Amoxicillin + Clavulanic Acid: 3984 patients; CIP: 5124 patients; Meropenem: 5190 patients; Tazobactam: 4402 patients | I | NA | Yes / Internal | No | 21/125 |
| Chan A-L, 2006 [32] | RCS | Taiwan | 1 | NA | To explore using simple, effective supervised learning techniques to predict vancomycin dosage | Patients who received a 5-days course of vancomycin | 833 | I | NA | Yes / Internal | No | 8 |
| Corbin C-K, 2020 [72] | RCS | USA | 1 | 2008–2014 | To develop ML models to predict bacterial susceptibility to antibiotics and create personalized antibiotics for patients, improving the empirical selection of antibiotics | Hospitalized patients with confirmed bacterial infections from microbial cultures | NA | I | NA | Yes / Internal | NA | 4261 |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|-------------------------|------------|-------------|------|---------------------|---|---|---|----|-----------------------------|----------------|--------|----------------------------|
| Corbin C-K, 2022 [20] | RCS | USA | 5 | Jan 2009 – Dec 2019 | To investigate the utility of ML-based clinical decision support for antibiotic prescribing stewardship | Patients who presented to Stanford ED, Massachusetts General Hospital, and Brigham and Women's Hospital in Boston | Stanford: 8342 infections from 6920 patients; Boston: 15806 uncomplicated UTI from 13862 patients. Training (5804 from 2009–2017), and validation (1218 from 2018), and test (1320 from 2019) | I | Stanford: NA; Boston: UTI | Yes / Internal | Yes | Stanford 43220; Boston 788 |
| de Vries S, 2022 [12] | RCS | Netherlands | 1 | Jan 2017–Dec 2018 | To report design and evaluation of a CDSS to predict UTI before the urine culture results are available | Inpatients of the University MC Utrecht | 906 cultures from 810 patients | I | UTI | Yes / External | No | 36 |
| Eickelberg G, 2020 [21] | RCS | Israel | 1 | 2011–2012 | To identify ICU patients with low risk of bacterial infection as candidates for earlier EAT discontinuation | ICU adults with suspected community-acquired bacterial infection | 10290 (12232 ICU encounters); split into training and test set following a 70/30 split | I | NA | Yes / Internal | No | NA |
| Feretzakis G, 2020 [92] | RCS | Greece | 1 | Jan 2017–Dec 2018 | To compare the performance of eight ML algorithms to assess antibiotic susceptibility predictions | ICU patients in a public tertiary hospital | 345 | I | Respiratory, urinary, wound | Yes / Internal | No | 23067 |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|-------------------------|------------|---------|------|-------------------|---|--|-----------------------|----|------------------------------------|----------------|--------|----------|
| Feretzakis G, 2020 [87] | OBS | Greece | 1 | Jun 2018-Dec 2018 | To evaluate the ability of different ML algorithms to predict antimicrobial resistance using patient demographic data, sample type, and Gram stain results | Patients hospitalized in two Internal Medicine Departments at a Greek public hospital | 5590 | I | NA | Yes / Internal | NA | 5 |
| Feretzakis G, 2021 [16] | RCS | Greece | 1 | 2018 | To assess the effectiveness of AutoML trained models to predict AMR | Patients admitted to the internal medicine wards of a public hospital | 499 (11496 instances) | I | NA | Yes / Internal | No | 6 |
| Feretzakis G, 2021 [88] | OBS | Greece | 1 | 2019 | To evaluate the performance of different ML models in predicting AMR for acinetobacter baumannii, K. pneumoniae, and P. aeruginosa using microbiological data | Patients hospitalized in the ICU of a Greek public hospital with isolates of acinetobacter baumannii, K. pneumoniae, and P. aeruginosa | 6086 | I | Respiratory, blood-stream, urinary | NA / NA | NA | NA |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|-----------------------------|------------|---------|------|--------------------|--|--|--|----|---|----------------|--------|----------|
| | | | | | | | | I | Bloodstream | Yes / Internal | No | |
| García-Vidal C, 2021 [65] | RCS | Spain | 1 | Jan 2008-Dec 2017 | Assess MDR-GNB infection risk factors via large data analysis and evaluate ML's ability to predict infection risk at febrile neutropenia onset | Hematological patients with febrile neutropenia | 349 (3235 episodes of febrile neutropenia) | | | Yes / Internal | No | 28/43 |
| Goodman K-E, 2022 [22] | RCS | USA | 1 | July 2017-Dec 2019 | To identify patient and treatment characteristics linked to intervention likelihood in a PAF program and develop models to safely exclude certain antimicrobial orders from review | Patient with antimicrobial orders from University of Maryland MC | 17503; testing set 3435 | I | Bacteremia, bone/joint, central nervous system, cardiac/vascular, etc | Yes / Internal | No | 33 |
| Guerrero-López A, 2023 [34] | RCS | Spain | 20 | 2014–2019 | To evaluate ML application in MALDI-TOF mass spectrometry for predicting K. pneumoniae resistance | Patients with K. pneumoniae infection | 402 | I | NA | Yes / Internal | No | 76 |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|--------------------------------|------------|-----------|----------|---|--|---|--|----|----------------|----------------|----------------|--|
| | | | | | | | | | | Yes / Internal | Yes / Internal | |
| Hassan N, 2023 [40] | RCS | UK | Multiple | 2023 | To develop and validate an AI model predicting post-operative infection in surgical patients | Surgical patients undergoing elective abdominal surgery | 2716 | I | Abdominal | Yes / Internal | No | 74 |
| Herman B, 2021 [42] | XS | Indonesia | Multiple | Model Building Jan 2015-Dec 2019; Testing Jan 2020-Oct 2020 | To develop and evaluate CUHAS-ROBUST, an AI tool for screening RR-TB in resource-limited settings lacking rapid diagnostic tests | Patients with suspected RR-TB | 487 for model building + 157 for testing (644 total) | I | Respiratory | Yes / Internal | No | 19 |
| Hernández-Carnero A, 2023 [49] | RCS | Spain | 1 | Jul 2004 -May 2019 | To develop a method using LSTM neural networks for predicting antibiotic resistance in pseudomonas aeruginosa nosocomial infections in the ICU | ICU patients from the University Hospital of Fuenlabrada with pseudomonas aeruginosa detected in cultures | Cultures: 755 AMG, 643 CAR, 749 Cephalosporins-4th generation, 749 Penicil-lins + β -lactamase inhibitors, 483 POLY, 708 QUI | I | NA | Yes / Internal | No | AMG: 36, CAR: 18, Cephalo-sporins-4th gen: 67, Penicil-lins + β -lactamase inhibitors: 24, POLY: 55, QUI: 49 |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|----------------------|------------|-------------|------|-------------------|--|---|--|-----|---|----------------|--------|----------|
| | | | | | | | | I | NA | Yes / Internal | No | 28/28 |
| Hirano Y, 2021 [61] | RCS | USA | 1 | 2008–2019 | To develop and validate an XGB ML model to predict MRSA in mechanically ventilated patients using MIMIC-IV data | Adult patients on mechanical ventilation screened for MRSA during their ICU stay | 809 screened for MRSA | | NA | Yes / Internal | No | 28/28 |
| Huang T-S, 2020 [39] | CCS | Taiwan | 1 | Jan 2016–Oct 2017 | Detection of CRKP using MALDI | NA | K. Pneumoniae: 46 resistant, 49 susceptible | NA | NA | Yes / Internal | No | NA |
| Ilhanlı N, 2024 [76] | RCS | South Korea | 1 | Oct 2012–Oct 2022 | To predict AMR in patients with a UTI using ML models and to interpret these models | Patients with UTI (19–100 years) | 3865 (Cephalexin 3535, TZP 737, TMP-SMX 708, Fluoroquinolone 1582, CAR 1365) | I/O | UTI | Yes / Internal | No | 71 |
| Jeon K, 2022 [70] | OBS | South Korea | 1 | 2018 | To evaluate the performance of a ML-based system for the presumptive identification of MRSA using MALDI-TOF and comparison of accuracy according to SCCmec types | S. aureus clinical isolates from various specimens (blood, sputum, ear swab, soft tissue, urine, tracheal aspiration, bronchoalveolar lavage, etc.) | 194 clinical isolates (106 MRSA and 88 MSSA) | NA | Bloodstream, soft tissue, urinary, respiratory, gastrointestinal, etc | Yes / Internal | Yes | NA |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|----------------------|------------|-------------|------|-------------------|--|--|--|----|----------------|----------------|--------|---|
| Jiménez F, 2020 [54] | ERS | Spain | 1 | Jan 2009-Jan 2018 | To develop and evaluate ML models, focusing on multivariate time series forecasting with feature selection, to predict antibiotic resistance outbreaks | Patients at risk of or with antibiotic-resistant infections, particularly in hospitals | NA | I | NA | Yes / Internal | No | Initially: 508; SVM: 13, DT: 7, RF: 5 |
| Kim C, 2023 [86] | CS | South Korea | 1 | 2010–2021 | To develop prediction models for antibiotic non-susceptibility to support the selection of empiric antibiotics for HA-UTIs | Patients aged (≥ 18 years) with suspected HA-UTIs | 10474 | I | UTI | Yes / Internal | Yes | Approximately 20000 predictors; final stage: 140 |
| Kong P-H, 2022 [50] | RCS | Taiwan | 1 | Dec 2020-Jul 2021 | To develop a ML model using MALDI-TOF data to discriminate MRSA from MSSA in patients with bacteremia | Patients with <i>S. aureus</i> bacteremia | 366 samples (147 MRSA, 219 MSSA) for training; 182 samples (72 MRSA, 110 MSSA) for testing | I | Bloodstream | Yes / Internal | No | Initially: 508; Included: SVM: 13, DT: 7, RF: 5, PR: 15 |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|----------------------------|------------|-----------|------|-------------------|---|---|--|----|-----------------------------------|----------------|--------|------------------------------|
| Lapp Z, 2021 [69] | PCS | USA | 21 | NA | Determine the importance of patient characteristics and CRKP genetic background in predicting infection risk in CRKP-colonized patients | Patients with extraintestinal CRKP colonization | 331 CRKP Sequence Type 258 isolates analyzed (355 isolates total, 24 were excluded for not being ST258) | I | Respiratory, urinary, bloodstream | Yes / Internal | No | 141 (50 patient, 91 genomic) |
| Lee A-L-H, 2021 [37] | RCS | Hong Kong | 3 | Jan 2015-Dec 2019 | To predict ESBL production in community-onset Enterobacteriaceae bacteraemia | Patients with community-onset bacteraemia | 5625; split into a training set and a test set in a 4:1 ratio | I | NA | Yes / Internal | No | 133 |
| Lewin-Epstein O, 2021 [73] | CS | Israel | 1 | May 2013-Dec 2015 | To predict the antibiotic resistance profiles of bacterial infections in hospitalized patients using ML algorithms applied to EHR | Hospitalized patients with culture-confirmed bacterial infections, covering various species and sites | More than 16000 AMR tests (samples: Ceftazidime: 2942, GEN: 4360, Imipenem: 2235, Ofloxacin: 3117, TMP-SMX: 3544) | I | NA | Yes / Internal | NA | NA |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|----------------------|------------|---------|------|-------------------|--|--|---|----|------------------------------------|----------------|--------|----------|
| | | | | | | | | I | Respiratory | Yes / External | No | 16 |
| Liang Q-Q, 2022 [51] | RCS | China | 2 | 2015–2019 | To develop and prospectively validate an ML model predicting CAR-GNB carriage within one week in ICU patients | ICU patients with potential CAR-GNB carriage | 10247; 1385 with positive CAR-GNB cultures, 1535 with negative CAR-GNB cultures | | | | | |
| Liang Q-Q, 2024 [77] | RCS | China | 1 | Jan 2015–Dec 2021 | To predict whether the pathogen causing bloodstream infections in the ICU is CAR-GNB using ML algorithms | Critically ill patients admitted to the ICU with suspected bloodstream infections | 952 positive blood cultures (CAR-GNB 418, non-CAR-GNB 534), 1422 negative blood; 70% training, 15% validation, 15% test | I | Bloodstream | Yes / Internal | NA | NA |
| Liu Q, 2024 [46] | RCS | China | 1 | 2012–2022 | To develop ML models predicting pathogens in secondary immunodeficiency patients, aiding antibiotic decisions before culture results | Patients with secondary immunodeficiency from conditions like hematological malignancies, autoimmune diseases, etc | 443 | I | Bloodstream, abdominal cavity, etc | Yes / Internal | No | 88 |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|------------------------------|------------|---------|------|-----------------|---|---|---|----|----------------|----------------|--------|--------------|
| Mancini A, 2020 [18] | RCS | Italy | 1 | 2012–2019 | To design, develop, and evaluate, with a real AS dataset, a predictive model useful for predicting MDR UTI onset after patient hospitalization | Patients with nosocomial UTI from Piemonte Hospital in Senigallia | 1486; randomly divided in training set (70%) and test set (30%) | I | UTI | Yes / Internal | Yes | 6 |
| Martínez-Agüero S, 2019 [64] | RCS | Spain | 1 | 2003–2015 | Develop ML models to predict antimicrobial resistance in <i>P. aeruginosa</i> , aiming for early detection of resistant bacteria to improve patient outcomes and limit the spread of infections | ICU patients | Total patients 2630; AMG: 2177; CAR: 1458; 4th-gen Cephalosporins: 1582; Broad-spectrum antibiotics: 2309; POLY: 570; QUI: 1952 | I | Bloodstream | Yes / NA | NA | 20–30/78–127 |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|------------------------------|------------|-----------|----------|-------------------|--|--|--|-----|--|----------------|--------|----------|
| Martínez-Agüero S, 2022 [57] | RCS | Spain | 1 | 2004–2020 | To develop interpretable models for early prediction of AMR in ICU patients using multivariate time-series data and intelligent feature selection | ICU patients with bacterial infections | 3470 (627 identified with AMR) | I | Respiratory, bloodstream, urinary, surgical sites, etc | Yes / NA | NA | 26/1252 |
| McFadden B-R, 2023 [63] | RCS | Australia | Multiple | Jan 2018-May 2020 | Establish an ML pipeline to predict blood culture outcomes from routine blood data, reducing unnecessary cultures and improving diagnostic stewardship | Adult patients whose blood samples were analyzed by Sysmex XN-2000 analyzers | Training: 10965 samples; internal validation: 318 samples; external validation: 1245 samples | I/O | Bloodstream | Yes / Internal | No | NA |
| McGuire R-J, 2021 [30] | CCS | USA | 1 | Jan 2012-Oct 2017 | To develop a prediction model for CAR resistance | All patients 18 years of age and older admitted to Barnes-Jewish Hospital / Washington University in St. Louis | 58752 patients with non-CAR resistant; 1088 patients with CAR resistant; randomly split into training (80%) and test (20%) data sets | I | NA | Yes / Internal | Yes | 67 |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|-------------------------|------------|----------|------|-------------------|---|---|--|----|----------------|----------------|--------|----------|
| Moehring R-W, 2021 [14] | RCS | USA | 3 | Oct 2015-Sep 2017 | To evaluate whether variables derived from the EHR accurately identify inpatient antimicrobial use | Adult and pediatric inpatient from Duke University Health System | 170294; 80% training and 20% testing sets randomly sampled | I | NA | Yes / Internal | Yes | 204 |
| Nigo M, 2024 [36] | RCS | USA | 2 | Jan 2018-Apr 2021 | To make an accurate risk stratification of MRSA | All patients from Memorial Hermann Hospital, Houston | Test 30557; validation 393713 | I | NA | Yes / External | No | NA |
| Oonsivilai M, 2018 [24] | RCS | Cambodia | 1 | Feb 2013-Jan 2016 | To predict Gram stains and whether bacterial pathogens could be treated with standard empiric antibiotic regimens | Children with at least one positive blood culture from Angkor Hospital for Children | AMP + GEN: 243; Ceftriaxone: 68; | I | Bloodstream | Yes / Internal | Yes | 35 |
| Page P-S, 2022 [89] | RCS | USA | 1 | 1995–2020 | To develop a predictive model to evaluate the likelihood of failure of medical therapy in the setting of SEA | All patients presenting with SEA from primary spinal infections | 159 | I | SEA | No / NA | Yes | 18 |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|------------------------------|------------|---------|------|-------------------|---|---|-----------------------------|----|----------------|----------------|--------|--|
| Pascual-Sanchez L, 2021 [74] | RCS | Spain | 1 | 2004–2020 | To anticipate the antibiotic outcomes associated with MR bacteria in ICU patients by applying ML techniques | ICU patients with MR bacteria | 3476 (628 with MR bacteria) | I | NA | Yes / Internal | NA | 174 initially, reduced through feature selection methods |
| Ravkin H-D, 2024 [68] | RCS | Israel | 1 | Jan 2010-Dec 2020 | Develop an ML tool to identify patients in ESBL-endemic areas who can be safely treated without ESBL-targeted therapy, reducing unnecessary broad-spectrum antibiotic use | Hospitalized patients with positive urine cultures for E. coli or K. pneumoniae | 17913 | I | UTI | Yes / Internal | Yes | 7 |
| Rhodes N-J, 2023 [67] | CCS | USA | 2 | Jan 2014-Mar 2018 | To develop and validate ML models to predict the risk of MRSA in hospitalized patients with community-acquired pneumonia | Adult patients (≥ 18 years) hospitalized with community-acquired pneumonia | 1823 | I | Respiratory | Yes / Internal | No | NA |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|-----------------------|------------|-------------|------|--------------------|---|--|---|-----|----------------|----------------|--------|----------|
| Rich S-N, 2022 [44] | RCS | USA | 47 | 2011–2019 | To develop a prediction model using EHR data to identify high-risk patients for antibiotic-resistant UTIs and guide antimicrobial therapy decisions | Patients diagnosed with a UTI who had antibiotic susceptibility tests | TMP-SMX: 9072; NIT: 9762; CIP: 9688; any MDR: 10340 | I/O | Urinary | Yes / External | Yes | NA |
| Schinke M, 2023 [38] | PCS | Netherlands | 1 | Oct. 2021–Sep 2022 | To predict blood culture outcomes and enhance diagnostic stewardship | Adult patients | 3035; testing 1606 | E | NA | Yes / Internal | No | NA |
| Sehult J-N, 2023 [41] | RCS | USA | 5 | Jan 2017–Dec 2017 | To develop and validate a ML DT algorithm using retrospective urinalysis and urine culture data | Adult patients from non-maternity inpatient and outpatient units | 38361 paired urinalysis and urine culture cases | I/O | Urinary | Yes / Internal | No | NA |
| Shang J-S, 2000 [90] | RCS | USA | 5 | Mar 1996–Mar 1997 | To investigate the potential of using NN and LR approach in diagnosing MRSA | S. aureus infected patients were obtained from five medical facilities in Pittsburgh | 504 | I | NA | Yes / Internal | No | 8 |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|------------------------|------------|---------|------|-------------------|---|--|---|----|---|----------------|--------|----------|
| Shi Z-Y, 2022 [13] | RCS | Taiwan | 25 | May 2013-May 2014 | To develop efficient ML models for auditing appropriate surgical antimicrobial prophylaxis | Patients with healthcare associated infections receiving at least one antimicrobial drug | 7377 | I | Bloodstream, urinary, respiratory and surgical site | Yes / Internal | No | 26 |
| Sophonsri A, 2023 [66] | RCS | USA | 2 | Mar 2014-Dec 2019 | Identify risk factors for development and mortality of vHABP and evaluate empiric antibiotic management with an ML approach | Hospitalized adults (≥ 18 years) who developed nosocomial pneumonia within the specified time frame | 457 (32.4% vHABP, 37.0% nvHABP, and 30.6% VABP) | I | NA | Yes / Internal | No | 18/50 |
| Stracy M, 2022 [19] | RCS | USA | 1 | 2008-2019 | To estimate patients' ICU LOS and mortality outcomes for any given day under the alternative scenarios of if they were to stop versus continue antibiotic treatment | Patients who received IV antibiotic treatment for a duration between 1-21 days during an ICU stay, at Beth Israel Deaconess MC, Boston | 18988 (22845 unique stays) | I | Respiratory, UTI | Yes / NA | No | 43 |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|-------------------------|------------|----------------------------|------|-------------------|---|---|---------------------------|----|------------------|----------------|--------|----------|
| Tacconelli E, 2020 [52] | PCS | Italy, Serbia, and Romania | 3 | Sep 2010-Jun 2013 | To measure the impact of antibiotic exposure on ESBL-GNB colonization, accounting for confounders using ML methods | Hospitalized patients in medical and surgical wards | 10034 (28322 rectal swab) | I | Gastrointestinal | Yes / NA | NA | 39 |
| Takács A-T, 2023 [48] | RCS | Hungary | 1 | Jan 2016-Dec 2017 | To develop a DT model using ML to distinguish EBV and CMV infections from other tonsillopharyngitis causes in children, reducing unnecessary antibiotic use | Children (0–18 years) with tonsillopharyngitis | 242 | I | Pharynx | Yes / Internal | No | 23 |
| Tran-The T, 2024 [47] | RCS | Republic of Korea | 1 | Jan 2014-Dec 2019 | To develop explainable ML models to assist ASP by prioritizing inpatients who would benefit most from interventions | Adult inpatients (18–90 years) receiving systemic antibiotics at the hospital | 19468 | I | NA | Yes / Internal | No | 406 |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|--------------------------|------------|---------|------|-------------------|---|---|--|----|----------------|----------------|--------|----------|
| Tsai W-C, 2023 [84] | RCS | Taiwan | 1 | Jan 2017-Dec 2020 | To evaluate the ability of the RF algorithm to predict bacteremia in adult febrile patients in the ED | Febrile adults (≥ 20 years) with at least one blood culture taken in the ED | 5647; divided into derivation (3369) and validation (2278) | E | NA | Yes / Internal | NA | 21 |
| Tsurumi A, 2023 [31] | RCS | USA | 7 | 2003–2009 | To enable precision medicine in pediatric burn care and develop predictive tools for early intervention in thermally injured children | Children with burns | 82 | E | Bloodstream | No / NA | No | NA |
| Tzelves L, 2022 [23] | RCS | Greece | 1 | 2019 | To evaluate the performance of ML techniques in predicting bacterial resistance in patients with urolithiasis | Patients with urolithiasis hospitalized in a urology department | 239 | I | UTI | Yes / Internal | NA | NA |
| Viswanathan V, 2024 [27] | PCS | India | 1 | NA | To evaluate the efficacy of auto-fluorescence imaging of diabetic foot ulcers using a novel artificial intelligence | Patient with diabetic foot ulcers | 178 (203 tissue samples) | O | Foot | No / NA | No | NA |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|---------------------|------------|---------|------|---|--|---|---|----|--|----------------|--------|----------------------------------|
| Wang C-X, 2022 [60] | RCS | Taiwan | 1 | Jun 2013-Feb 2018 | To develop and validate ML models for rapid detection of CIRKP using mass spectrometry data combined with patient demographics | Patients infected with <i>K. pneumoniae</i> | 16697 samples initially, with 15782 samples selected after quality control (11354 CISKP and 4428 CIRKP) | I | Bloodstream, body fluid, wound, urinary, etc | Yes / Internal | No | 102/480 |
| Wang J, 2022 [81] | CS | China | 1 | Jan 2020-Dec 2021 | To evaluate the ability of RF, SVM, and SVM with Kernel algorithms to distinguish CRKP from CSKP using MALDI-TOF data | Clinical isolates of <i>K. pneumoniae</i> (95 CRKP and 76 CSKP) | 171 isolates (95 CRKP, 76 CSKP) | I | NA | Yes / Internal | No | 211, with 47 house-keeping peaks |
| Wang Y, 2023 [29] | PCS | China | 1 | Aug 2021-Jan 2022; validation May 2022-Jul 2022 | To develop a model for predicting MDRO infection in critically ill patients | Patients admitted to a ICU | 688; divided randomly into training set (80%) and test set (20%); validation set 238 | I | NA | Yes / External | Yes | 9 |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|---------------------|------------|-----------|------|--|---|--|---|----|---|----------------|--------|----------|
| Wang Z, 2021 [59] | RCS | Taiwan | 2 | Discovery cohort 2013–2019; Replication cohort 2015–2017 | To build a predictive model using mass spectrometry and demographic data to characterize methicillin resistance in <i>S. aureus</i> | Patients with <i>S. aureus</i> infections | Discovery population: 23099; replication population: 4962 | I | Wound, respiratory, blood-stream, sterile body fluid, UTI | Yes / External | No | 217/217 |
| Wong J-G, 2020 [26] | PCS | Singapore | 1 | Jun 2016–Nov 2018 | To develop prediction models based on local clinical and laboratory data to guide antibiotic prescribing for adult patients with uncomplicated upper respiratory tract infections | Patients with uncomplicated URTI at the ED at Tan Tock Seng Hospital | 715; using 70% of the participants as training set | E | Respiratory | Yes / Internal | No | 50 |

Table 2 (continued)

| Author, year (Ref) | Study type | Country | Cen. | Data time frame | Study objective | Target population | Number of patients | HS | Infection site | TDS/Validation | Calib. | Features |
|---------------------|------------|---------|------|-------------------|---|--|---|-----|--|----------------|--------|----------|
| Yu J, 2023 [78] | RCS | Taiwan | 1 | Oct 2022-Dec 2022 | To evaluate the ability of a ML-based MALDI-TOF approach combined with the Rapid Sepsityper protocol to predict MRSA and CRKP from flagged blood cultures | Patients with monocrobial bloodstream infections | 461 (MRSA: 44 S. aureus isolates; CRKP: 126 K. pneumoniae isolates) | I | Bloodstream | Yes / Internal | NA | NA |
| Zhang Y-M, 2023 [2] | RCS | Taiwan | 1 | Apr 2018-Mar 2022 | To establish an artificial NN model based on MALDI-TOF MS data to rapidly and accurately identify CRKP from CSKP | Clinical isolates of K. pneumoniae collected from various specimens (urine, sputum, blood) | 2683 isolates (369 CRKP and 2314 CSKP) | I/O | Urinary, bloodstream, wound, vagina, etc | Yes / External | NA | 4500 |

RCS Retrospective Cohort Study; ERS Evaluative Research Study; OBS: Observational Study; PCS Prospective Cohort Study; CCS Case-Control Study; XS Experimental Study; AMR Antibiotic Resistance; ICU Intensive Care Unit; EHR Electronic Health Record; MALDI-TOF Matrix-Assisted Laser Desorption/Ionization Time-of-Flight; ML Machine Learning; NLP Natural Language Processing; NA Not Applicable; MDRO Multidrug-Resistant Organism; CIRKP Carbapenem-Resistant Klebsiella pneumoniae; MRSA Methicillin-Resistant Staphylococcus aureus; CSKP Carbapenem-Susceptible Klebsiella pneumoniae; VRE Vancomycin-Resistant Enterococcus; VSE Vancomycin-Susceptible Enterococcus; TMP-SMX Trimethoprim-Sulfamethoxazole; CIP Ciprofloxacin; EAT Empirical Antibiotic Therapy; CCT Clinical Decision Support Tool; HA-UTI Hospital-Acquired Urinary Tract Infection; S. Staphylococcus; GNB Gram-Negative Bacteria; Bacteremia Presence of bacteria in the blood; LOS Length of Stay; SEA Spinal Epidural Abscess; RR-TB Rifampicin-Resistant Tuberculosis; vHABP Ventilator-Associated Hospital-Acquired Bacterial Pneumonia; CRKP Carbapenem-Resistant Klebsiella pneumoniae; NIT Nitrofurantoin

internal and external. The number of features used in the studies varied greatly from a minimum of 5 [92] features to a maximum of 43,220 [20], with a mean of 1,375.3. Calibration was performed only in 16 models [14, 17, 18, 20, 24, 29, 30, 33, 43–45, 68, 70, 71, 86, 89].

Characteristics of the features of the included studies

Figure 2 illustrates the distribution of key features analysed in the included studies. The most common were microbiological/laboratory data (91.3%, $n=73$) [12, 14–26, 28–43, 46–60, 62–70, 72, 73, 75–90, 92], followed by clinical data (82.5%, $n=66$) [12–15, 17, 19–26, 28–31, 33, 36–38, 40–62, 64–69, 71–78, 82, 84–90]. Only 11.3% ($n=9$) of the studies included paediatric populations [12–14, 24, 31, 41, 48, 75, 82].

ML algorithms and performance

Table 3 summarizes the algorithms used in the included studies. The most used algorithm was RF ($n=39$) [12, 14, 16, 20–22, 24, 28, 35, 39, 42, 44–46, 50–52, 54, 60, 62–66, 69, 72, 74–77, 80–82, 84–87, 91, 92]. Boosted models were applied 34 times, with XGB used in 21 cases [12, 21, 28, 30, 35, 42, 47, 51, 53, 55, 56, 60, 61, 63, 65, 72, 74, 77, 82, 84, 86], along with other Boosting algorithms (Boosted DT $n=3$ [17, 24, 32], Gradient Boosted DT $n=3$ [20, 68, 73], GBM $n=3$ [43, 46, 65], Boosted LR $n=2$ [44, 46], CatBoost $n=1$ [18], AdaBoost ($n=1$) [13]). SVM models were used 22 times (most frequently SVM $n=13$ [12, 18, 21, 34, 39, 50, 54, 58, 60, 77, 80, 82, 85]) and SVM with RBF kernel $n=4$ [24, 34, 69, 81]). LASSO was used 8 times [20, 26, 31, 59, 72–74, 86], primarily for variable selection and dimensionality

reduction. Neural Networks, including both general NN (Artificial NN $n=5$ [33, 42, 54, 58, 83], NN $n=4$ [18, 37, 73, 90], Backpropagation NN $n=1$ [29], NN with SHAP $n=1$ [71]) and more advanced variants like Recurrent NNs (LSTM $n=2$ [49, 57], Bidirectional LSTM $n=2$ [19, 57], Recurrent NN $n=2$ [25, 36], GRU $n=1$ [57]), were also widely employed, totalling 18 instances. DT models were also common, with DT used in 16 cases [13, 24, 28, 32, 41, 42, 44, 48, 50–52, 58, 64, 67, 74, 77], J48 (C4.5) in three [85, 87, 92], and CART in two [26, 75].

Moreover, Table 3 provides a comprehensive quantitative summary of the predictive performance of various ML models, as measured by AUC. The AUC values range from 41.6 [49] to 99.2 [13]. It was evaluated in three main contexts: predicting antibiotic resistance using ML models, assessing resistance to specific antibiotics, and predicting pathogen-specific resistance to various antibiotics.

Accuracy ranged from 41.6 [49] to 100.0 [75], the latter achieved by an RF model (Table S2). Sensitivity values varied from 0.0 [49], for predicting Quinolone resistance by an LSTM model to 100.0 [63, 82], attained by RF and XGB models (Table S3). Specificity ranged from 15.0 [63] to 100.0 [39, 49, 65], with RF, Naïve Bayes, and LSTM models reaching the upper limit (Table S4). PPV ranged from 6.3 [67] to 100.0 [65, 71, 82]; NPV spanned from 5.0 [71] to 99.6 [67] (Table S5).

Meta-analysis

In our meta-analysis, ML models demonstrated strong predictive performance and diagnostic accuracy across various AMS settings, despite the considerable heterogeneity observed among the included studies. For the AUC, 25 studies and 95 ML models were analyzed. The fixed

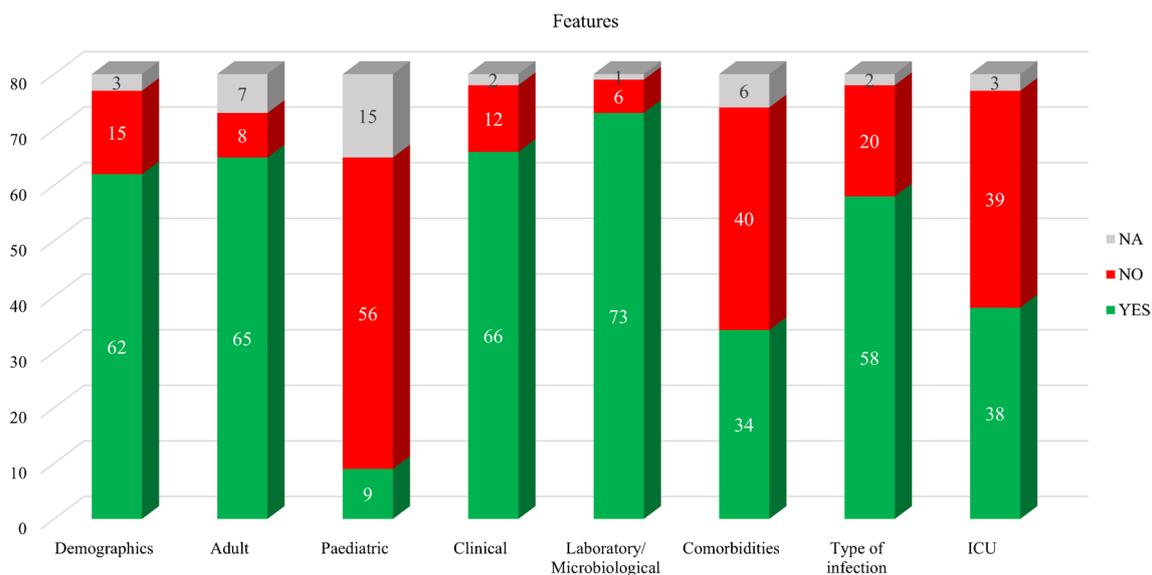


Fig. 2 Distribution of the main features included in the Machine Learning (ML) models. NA = not available; ICU = Intensive care Unit setting

Table 3 Machine Learning (ML) Algorithms and Area Under the Curve (AUC) of ML models from the included studies

| Author, year | ML Algorithms | % Area Under the Curve |
|------------------------|---|--|
| Abu-Aqil G, 2022 | SVM, XGB, RF, k-NN | Amoxicillin: LDA 67.0, XGB 78.0, RF 76.0; Cefuroxime axetil: LDA 75.0, XGB 83.0, RF 82.0; GEN: LDA 73.0, XGB 83.0, RF 83.0; Piperacillin: LDA 67.0, XGB 75.0, RF 72.0; SMX: LDA 68.0, XGB 80.0, RF 79.0 |
| Abu-Aqil G, 2023 | MLP, DT, SimpleLogistic, and DT with SpreadSubsample, Bagging, SMOTE and AdaBoost | Ceftazidime: 86.0; Ceftriaxone: 85.0; Cefuroxime: 83.0; Cephalixin: 84.0; CIP: 89.0 |
| Abu-Aqil G, 2023 | A 2-stage RF | Ceftazidime: proteus: 80.0, P. aeruginosa: 78.0; CIP: proteus: 86.0, P. aeruginosa: 75.0; GEN: proteus: 82.0, P. aeruginosa: 85.0 |
| Abu-Aqil G, 2024 | Combined system, Learning modules | Taxonomic classification of K. pneumoniae versus other UTI bacteria: 99.0; Amoxicillin: 77.0; Ceftazidime: 78.0; Cephalixin: 78.0; CIP: 79.0; Piperacillin: 74.0 |
| Agbaria A-H, 2020 | Ensemble Model combining Algorithms, VotingEnsemble, SparseNormalizer with XGB and LightGBM | First Step (Control vs Infected): Naïve Bayes: 97.0, XGB: 98.0, RF: 97.0, SVM: 96.0; Second Step (Viral vs Bacterial): Naïve Bayes: 91.0, XGB: 93.0, RF: 93.0, SVM: 91.0 |
| Al-khelifeh E-M, 2024 | Boosted DT | NA |
| Ananda-Rajah M-R, 2017 | Catboost, SVM, NN | Baseline Text Classifier: 73.9 [95% CI 67.1–80.6]; Naive Bayes: 92.8 [95% CI 88.0–97.5]; RF: 94.1 [95% CI 89.8–98.3]; SMO: 92.4 [95% CI 87.5–97.3]; J48(C4.5): 87.2 [95% CI 80.6–93.7] |
| Ayyildiz H, 2021 | Bidirectional LSTM (create with PyTorch) | NA |
| Beaudoin M, 2016 | RF, Gradient Boosted DT, LASSO, Ridge Regression | NA |
| Bhavani S, 2020 | RF, XGB, MLP, SVM, k-NN, VotingEnsemble | Bacteremia Prediction: 78.0 [95% CI 77.0–78.0] |
| Bolton W-J, 2022 | RF | 77.0 [95% CI 73.0–80.0] |
| Bolton W-J, 2024 | SVM C-support Vector; SMO; k-NN; J48(C4.5); RF; RIPPER; MLP | MIMIC—Short model: 1st threshold: 78.0 [sd ± 2.0], 2nd threshold: 69.0 [sd ± 3.0]; MIMIC—Long model: 1st threshold: 80.0 [sd ± 1.0], 2nd threshold: 75.0 [sd ± 2.0]; MIMIC Incomplete absorption—Short model: 1st threshold: 73.0 [sd ± 3.0], 2nd threshold: 67.0 [sd ± 5.0]; MIMIC Incomplete absorption—Long model: 1st threshold: 77.0 [sd ± 2.0], 2nd threshold: 73.0 [sd ± 3.0]; MIMIC Malabsorption—Short model: 1st threshold: 76.0 [sd ± 10.0], 2nd threshold: 75.0 [sd ± 11.0]; MIMIC Malabsorption—Long model: 1st threshold: 75.0 [sd ± 7.0], 2nd threshold: 71.0 [sd ± 16.0]; MIMIC UTI—Short model: 1st threshold: 77.0 [sd ± 3.0], 2nd threshold: 74.0 [sd ± 4.0]; MIMIC UTI—Long model: 1st threshold: 78.0 [sd ± 2.0], 2nd threshold: 77.0 [sd ± 4.0]; MIMIC Pneumonia—Short model: 1st threshold: 76.0 [sd ± 3.0], 2nd threshold: 76.0 [sd ± 3.0]; MIMIC Pneumonia—Long model: 1st threshold: 77.0 [sd ± 2.0], 2nd threshold: 74.0 [sd ± 4.0]; MIMIC Sepsis—Short model: 1st threshold: 82.0 [sd ± 5.0], 2nd threshold: 79.0 [sd ± 12.0]; MIMIC Sepsis—Long model: 1st threshold: 77.0 [sd ± 7.0], 2nd threshold: 76.0 [sd ± 18.0]; eICU—Short model: 1st threshold: 72.0 [sd ± 2.0], 2nd threshold: 65.0 [sd ± 5.0]; eICU—Long model: 1st threshold: 72.0 [sd ± 2.0], 2nd threshold: 64.0 [sd ± 6.0]; BEST MODEL 80.0 [sd ± 1.0] |
| Brokowski T-J, 2022 | RF, DT, Boosted DT, Linear SVM, Polynomial SVM, SVM with RBF kernel, kNN | LEV 90.0; Clindamycin 88.0; Vancomycin 87.0; Linezolid 93.0; Oxacillin 89.0; Gentamicin 91.0; Erythromycin 88.0; Daptomycin 92.0. Model not specified |
| Brown D-G, 2023 | Recurrent NN autoencoder | 10-Features RF Model: 67.0 [95% CI 66.0–68.0]; 4-Features RF Model: 63.0 [95% CI 62.0–64.0] |

Table 3 (continued)

| Author, year | ML Algorithms | % Area Under the Curve |
|----------------------|---|--|
| Bystritsky R-J, 2020 | LASSO, CART | 75.0 [95% CI 72.0–79.0] |
| Çağlayan Ç, 2022 | Illuminate® (Aduvuo Diagnostics Private Limited, Chennai, India), Algorithm not defined | RF VRE 77.0; XGB VRE 77.0; RF MRSA 70.0; XGB MRSA 66.0; RF CRE 72.0; XGB CRE 72.0; RF MDRO 89.0; XGB MDRO 87.0; DT MDRO 81.0 |
| Cai T, 2023 | RF, XGB, DT | 86.7 |
| Candela A, 2022 | Backpropagation NN | NA |
| Cavallaro M, 2023 | XGB | Amoxicillin + Clavulanic Acid: 78.0 [95% CI 76.0–81.0]; CIP: 87.0 [95% CI 85.0–89.0]; Meropenem: 99.0 [95% CI 98.0–99.0]; Tazobactam: 79.0 [95% CI 75.0–81.0] |
| Chan A-L, 2006 | LASSO | NA |
| Corbin C-K, 2020 | DT, Boosted DT | E. coli [N=2,424]: AMP: best model: RF 60.0 [95% CI 54.0–65.0]; Cefepime: best model: XGB 73.0 [95% CI 65.0–78.0]; Ceftriaxone: best model: RF 63.0 [95% CI 56.0–69.0]; Cefazolin: best model: RF 60.0 [95% CI 54.0–67.0]; CIP: best model: RF 67.0 [95% CI 63.0–72.0]; LEV: best model: RF 67.0 [95% CI 64.0–72.0]. K. pneumoniae [N=671]: Cefepime: best model: RF 74.0 [95% CI 62.0–84.0]; Ceftriaxone: best model: RF 70.0 [95% CI 61.0–78.0]; Cefazolin: best model: RF 61.0 [95% CI 52.0–70.0]; CIP: best model: LASSO 56.0 [95% CI 44.0–67.0]; LEV: best model: XGB 56.0 [95% CI 44.0–67.0]. P. aeruginosa [N=693]: Cefepime: best model: RF 66.0 [95% CI 50.0–76.0]; CIP: best model: XGB 66.0 [95% CI 53.0–75.0] |
| Corbin C-K, 2022 | Artificial NN | STANFORD DATASET Vancomycin Gradient Boosted DT 72.0 [95% CI 68.0–75.0]; AMP Gradient Boosted DT 62.0 [95% CI 59.0–65.0]; Cefazolin Gradient Boosted DT 67.0 [95% CI 64.0–70.0]; CIP RF 61.0 [95% CI 58.0–64.0]; Ceftriaxone Gradient Boosted DT 69.0 [95% CI 66.0–72.0]; Cefepime RF 65.0 [95% CI 61.0–69.0]; Vancomycin + Ceftriaxone Gradient Boosted DT 67.0 [95% CI 63.0–71.0]; Meropenem Gradient Boosted DT 69.0 [95% CI 65.0–72.0]; TZP RF 64.0 [95% CI 59.0–69.0]; Vancomycin + TZP RF 70.0 [95% CI 62.0–77.0]; Vancomycin + Cefepime RF 70.0 [95% CI 62.0–78.0]; Vancomycin + Meropenem Gradient Boosted DT 73.0 [95% CI 65.0–81.0] |
| de Vries S, 2022 | Gaussian Process with PIKE, KSSHIBA with RBF kernel, SVM with RBF kernel | BOSTON DATASET TMP-SMX Gradient Boosted DT 60.0 [95% CI 58.0–62.0]; NIT Gradient Boosted DT 57.0 [95% CI 54.0–61.0]; CIP LASSO 64.0 [95% CI 60.0–68.0]; LEV LASSO 64.0 [95% CI 60.0–67.0] SVM: 78.2 [sd ± 0.9]; XGB: 80.1 [sd ± 1.1]; RF 80.0 [sd ± 1.0]; k-NN: 78.6 [sd ± 1.0] |
| Eickelberg G, 2020 | LDA, XGB, RF | 72 h: RF: 79.3; XGB: 79.5; MLP: 77.9; SVM: 77.8; k-NN: 73.4; VotingEnsemble: 79.3; 48 h: RF: 78.8; XGB: 76.9; MLP: 77.1; SVM: 77.3; kNN: 73.3; VotingEnsemble: 78.8; 24 h: RF: 77.4; XGB: 77.6; MLP: 76.4; SVM: 76.3; k-NN: 71.4; VotingEnsemble: 77.0 |
| Feretakis G, 2020 | Recurrent NN | LibrarySVM C-Support Vector 66.0; SMO 65.9; 1-Nearest Neighbors 68.2; 5-Nearest Neighbors 71.1; J48(C4.5) 72.4; RF 70.3; RIPPER 69.9; MLP 72.6 |
| Feretakis G, 2020 | NN | Multinomial LR: 75.8; MLP: 74.6; k-NN: 72.7; J48 (C4.5): 72.2; RF: 72.1 |

Table 3 (continued)

| Author, year | ML Algorithms | % Area Under the Curve |
|---------------------------|--|---|
| Ferezakis G, 2021 | Blood culture prediction tool. Algorithm not specified | StackEnsemble 85.0; VotingEnsemble 85.0; SparseNormalizer, XGB 84.2; SparseNormalizer, LightGBM 83.7 |
| Ferezakis G, 2021 | RF, Naïve Bayes, k-NN, SVM | Original Data: JRip: 87.0, RF: 86.9, MLP: 86.5, Classification via Regression: 93.3, REPT: 87.3; Balanced Data: JRip: 85.7, RF: 87.2, MLP: 86.3, Classification via Regression: 91.8, REP: 87.1 |
| Garcia-Vidal C, 2021 | Ensemble Model combining Algorithms: using GBM and other ML algorithms | RF: 78.9; GBM: 78.7; XGB: 79.4 |
| Goodman K-E, 2022 | Pittsburgh Urinary Tract (PittUDT) DT | 76.0 [95% CI 75.0–77.0] |
| Guerrero-López A, 2023 | Artificial NN, DT, RF, XGB | K. pneumoniae wild type 75.0 [sd ± 11.0]; K. pneumoniae ESBL 72.0 [sd ± 14.0]; K. pneumoniae ESBL + CAR-producers 85.0 [sd ± 14.0] |
| Hassan N, 2023 | GBM | 88.6 in training; 85.5 in validation |
| Herman B, 2021 | Boosted LR, RF, DT | Artificial NN 96.0 in training data |
| Hernández-Carnero À, 2023 | RF | AMG: without feature selection: 76.0 [sd ± 5.7], with feature selection: 76.8 [sd ± 5.6]; CAR: without feature selection: 65.8 [sd ± 3.5], with feature selection: 66.5 [sd ± 3.3]; Cephalosporins—4th gen: without feature selection: 63.0 [sd ± 2.9], with feature selection: 63.5 [sd ± 2.6]; Penicillins + β-lactamase inhibitors: without feature selection: 71.1 [sd ± 3.9], with feature selection: 71.6 [sd ± 3.5]; POLY: without feature selection: 41.6 [sd ± 6.1], with feature selection: 42.3 [sd ± 6.7]; QUI: without feature selection: 88.3 [sd ± 3.7], with feature selection: 89.4 [sd ± 3.3] |
| Hirano Y, 2021 | k-NN, Boosted LR, RF, GBM | 89.0 [95% CI 83.0–95.0] |
| Huang T-S, 2020 | XGB, LightGBM | NA |
| İlhanlı N, 2024 | DT | RF—Training: Cephalosporin: 77.7 [95% CI 77.5–77.9]; TZP: 86.4 [95% CI 86.2–86.7]; CAR: 87.7 [95% CI 87.4–88.0]; TMP-SMX: 88.1 [95% CI 87.9–88.2]; Fluoroquinolone: 88.4 [95% CI 88.4–88.5]. RF—Test: Cephalosporin: 63.8 [95% CI 63.5–64.2]; TZP: 63.0 [95% CI 62.6–63.4]; CAR: 66.5 [95% CI 65.9–67.1]; TMP-SMX: 67.0 [95% CI 66.6–67.3]; Fluoroquinolone: 72.1 [95% CI 71.8–72.4] |
| Jeon K, 2022 | LSTM | total 87.6; blood isolates 90.6; non-blood isolates 86.3 |
| Jiménez F, 2020 | SVM, DT, RF, PR | SVM: 86.6; RF: 82.0; Artificial NN: 80.0; Gaussian Processes: 81.0 |

Table 3 (continued)

| Author, year | ML Algorithms | % Area Under the Curve |
|-------------------------|--|---|
| Kim C, 2023 | DT, RF, XGB | Full Model: AMP: LASSO 73.4, GBM: 72.4, RF: 72.3, stacked ensemble: 68.8; AMP/Sulbactam: LASSO: 71.0, GBM: 74.8, RF: 72.2, stacked ensemble: 76.9; Cefepime: LASSO: 66.0, GBM: 70.7, RF: 67.3, stacked ensemble: 76.1; CIP: LASSO: 71.7, GBM: 70.4, RF: 70.7, stacked ensemble: 77.0; GEN: LASSO: 62.0, GBM: 66.9, RF: 65.9, stacked ensemble: 64.0; Imipenem: LASSO: 61.8, GBM: 73.0, RF: 75.2, stacked ensemble: 80.6; TZP: LASSO: 69.5, GBM: 65.0, RF: 68.8, stacked ensemble: 76.1; TMP/SMX: LASSO: 73.9, GBM: 72.1, RF: 75.7, stacked ensemble: 76.5. Parsimonious Model: AMP: LASSO: 70.6, GBM: 70.2, RF: 69.1, stacked ensemble: 63.7; AMP/Sulbactam: LASSO: 71.8, GBM: 68.6, RF: 67.0, stacked ensemble: 55.9; Cefepime: LASSO: 73.0, GBM: 70.4, RF: 70.9, stacked ensemble: 57.4; CIP: LASSO: 70.0, GBM: 70.3, RF: 69.8, stacked ensemble: 62.5; GEN: LASSO: 65.9, GBM: 62.3, RF: 59.4, stacked ensemble: 54.0; Imipenem: LASSO: 70.8, GBM: 70.7, RF: 68.3, stacked ensemble: 71.3; TZP: LASSO: 64.4, GBM: 67.2, RF: 63.9, stacked ensemble: 56.0; TMP/SMX: LASSO: 73.0, GBM: 71.0, RF: 68.5, stacked ensemble: 54.5 SVM: 86.6; DT: 79.6; RF: 82.4; PR: 82.4 Median range = 55.0–68.0 across different feature sets and models 76.1 [95% CI 72.5–79.7] Ensemble models: Bacterial species excluded: Imipenem 79.2, Ofloxacin 76.3, Cefazidime 76.0, SMX-TMP 74.3, Combined 73.9, GEN 72.7; Bacterial species included: Imipenem 88.1, Ofloxacin 80.3, Cefazidime 82.0, SMX-TMP 79.9, Combined 80.8, GEN 80.4. Ensemble antibiotics performance: Bacterial species excluded: LASSO 75.3, XGB 74.8, NN 74.3, Ensemble models 75.7; Bacterial species included: LASSO 82.0, XGB 82.0, NN 80.3, Ensemble models 82.1 Validation set: RF 91.0, XGB 91.0, DT 90.0; Test set: RF 90.0, XGB 89.0, DT 89.0 Bloodstream Infection Model: DT: 77.0, RF: 86.0, SVM: 83.0, XGB: 85.0; CR-GNB Bacteremia Model: DT: 69.0, RF: 87.0, SVM: 88.0, XGB: 80.0 k-NN: Micro-average: 87.0, Macro-average: 91.0; Boosted LR: Micro-average: 93.0, Macro-average: 92.0; RF: Micro-average: 98.0, Macro-average: 98.0; GBM: Micro-average: 98.0, Macro-average: 97.0 Catboost 73.9 [sd±2.1]; SVM 62.8 [sd±2.5]; NN 65.2 [sd±2.3] NA GRU: 66.0 [sd±1.3]; LSTM: 66.7 [sd±1.8]; Bidirectional LSTM: 64.4 [sd±0.8] Internal validation set: XGB 76.0, RF 82.0; External validation set: 76.0 Detection CAR resistant 84.6 Adult all antibacterials 87.4 [95% CI 87.0–87.8]; Adult C. difficile agents 86.8 [95% CI 86.4–87.3]; Adult resistant Gram + 91.0 [95% CI 90.6–91.3]; Adult narrow spectrum beta-lactam 88.7 [95% CI 88.3–89.2]; Pediatric antibacterials 93.3 [95% CI 92.6–93.9]; Pediatric azithromycin 94.4 [95% CI 92.6–96.3]; Pediatric C. difficile agents 94.2 [95% CI 93.4–94.9]; Pediatric Gram + 94.6 [95% CI 93.8–95.5] |
| Kong P-H, 2022 | RF, DT | |
| Lapp Z, 2021 | XGB | |
| Lee A-L-H, 2021 | Gaussian Processes, Artificial NN, RF, SVM | |
| Lewin-Epstein O, 2021 | XGB | |
| Liang Q-Q, 2022 | XGB | |
| Liang Q-Q, 2024 | MLP, GRU, LSTM, Bidirectional LSTM (with Full Feature, Balanced Binary Cross-Entropy, Masking) | |
| Liu Q, 2024 | k-NN, Artificial NN, SVM, DT | |
| Mancini A, 2020 | LASSO | |
| Martínez-Agüero S, 2019 | SVM, XGB, RF | |
| Martínez-Agüero S, 2022 | XGB | |
| McFadden B-R, 2023 | LightGBM, RF | |
| McGuire R-J, 2021 | XGB, RF | |
| Moehring R-W, 2021 | k-NN, DT, RF, MLP | |

Table 3 (continued)

| Author, year | ML Algorithms | % Area Under the Curve |
|-------------------------|---|--|
| Nigo M, 2024 | RF, GBM, XGB | 91.1 [95% CI 90.0–91.6] |
| Oonsivilai M, 2018 | RF | Resistant to ceftriaxone: RF: 80.0 [95% CI 66.0–94.0]; Boosted DT: 88.0 [95% CI 77.0–100.0]; DT: 79.0 [95% CI 65.0–92.0]; Regularized LR: 75.0 [95% CI 58.0–91.0]; k-NN: 59.0 [95% CI 42.0–77.0]; SVM Radial Kernel: 80.0 [95% CI 64.0–96.0]; SVM Linear Kernel: 81.0 [95% CI 68.0–95.0]; SVM Polynomial Kernel: 83.0 [95% CI 71.0–95.0] |
| Page P-S, 2022 | DT(Classification Tree Analysis) | Resistant to AMP + GEN: RF: 74.0 [95% CI 59.0–89.0]; Boosted DT: 61.0 [95% CI 43.0–78.0]; DT: 54.0 [95% CI 46.0–63.0]; Regularized LR: 59.0 [95% CI 43.0–75.0]; k-NN: 60.0 [95% CI 43.0–76.0]; SVM Radial Kernel: 56.0 [95% CI 38.0–73.0]; SVM Linear Kernel: 58.0 [95% CI 41.0–75.0]; SVM Polynomial Kernel: 63.0 [95% CI 46.0–80.0] |
| Pascual-Sanchez L, 2021 | Gradient Boosted DT | Gram stain: RF: 71.0 [95% CI 57.0–86.0]; Boosted DT: 62.0 [95% CI 46.0–79.0]; DT: 63.0 [95% CI 48.0–77.0]; Regularized LR: 66.0 [95% CI 51.0–82.0]; k-NN: 61.0 [95% CI 45.0–77.0]; SVM Radial Kernel: 68.0 [95% CI 53.0–84.0]; SVM Linear Kernel: 75.0 [95% CI 60.0–89.0]; SVM Polynomial Kernel: 75.0 [95% CI 62.0–89.0] |
| Ravkin H-D, 2024 | Ridge regression, Elastic Net, RF, SVM with RBF kernel | 80.4 |
| Rhodes N-J, 2023 | Enhanced RF | Experiment 1 (Statistics of all features): highest: 76.0 with XGB and Chi-Square feature selection; Experiment 4 (Raw data): highest: 85.0 with XGB and LASSO feature selection; Experiment 2 (Patient features only): highest: 71.0 with XGB and Chi-Square; Experiment 3 (ICU features only): highest: 72.0 with RF and Chi-Square |
| Rich S-N, 2022 | NN with SHAP | 72.0 |
| Schinkel M, 2023 | LASSO, RF, XGB | 77.5 |
| Seheult J-N, 2023 | LASSO, Gradient Boosted DT, NN, Ensemble Model combining Algorithms | Resistant UTI: TMP-SMX: Boosted LR: 58.0 [SD 1.0], DT: 57.0 [SD 1.0], RF: 57.0 [SD 1.0]; NIT: Boosted LR: 62.0 [SD 1.0], DT: 60.0 [SD 2.0], RF: 57.0 [SD 1.0]; CIP: Boosted LR: 64.0 [SD 1.0], DT: 62.0 [SD 1.0], RF: 63.0 [SD 1.0]. MDR UTI: Boosted LR: 66.0 [SD 2.0], DT: 63.0 [SD 2.0], RF: 62.0 [SD 3.0] |
| Shang J-S, 2000 | RF, LASSO, XGB, DT, MLP | 78.0 [sd±2.0] |
| Shi Z-Y, 2022 | RF, CART | Urine WBC: 79.0; Leukocyte Esterase: 78.0; Bacteria: 77.0 |
| Sophonsri A, 2023 | RF | NN 92.8 [sd±1.6] DT with SpreadSubsample: 97.8; DT: 98.5; Bagging with DT: 99.1; Adaboost with DT: 98.3; DT with SMOTE: 98.5; SimpleLogistic: 98.7; MLP: 99.2 vHABP: 78.0 [95% CI 70.0–85.0]; nvHABP: 80.0 [95% CI 71.0–89.0]; VABP: 83.0 [95% CI 75.0–90.0] |
| Stracy M, 2022 | RF, SVM, DT, XGB | UTI: TMP-SMX 66.0; CIP 74.0; Amoxicillin/Clavulanic Acid 62.0; Cefuroxime—Axetil 67.0; Cephalexin 70.0; NIT 89.0; Fosfomycin 74.0. Wound Infections: TMP-SMX 63.0; CIP 60.0; Amoxicillin/CA 96.0; Cefuroxime—Axetil 58.0; Cephalexin 68.0 |
| Tacconelli E, 2020 | LightGBM | Validation set: RF 91.8, XGB 91.2, DT 90.7; Test set: RF 90.8, XGB 89.4, DT 89.8 |

Table 3 (continued)

| Author, year | ML Algorithms | % Area Under the Curve |
|---------------------|---|---|
| Takács A-T, 2023 | RF | GPT as a standalone marker: 86.0 |
| Tran-The T, 2024 | SVM, RF, Partial Least Squares-Discriminant Analysis | Discontinuation: XGB: 80.0, LightGBM: 80.0; IV to Oral: XGB: 80.0, LightGBM: 81.0; Early De-escalation: XGB: 78.0, LightGBM: 78.0; Late De-escalation: XGB: 69.0, LightGBM: 72.0 |
| Tsai W-C, 2023 | RF, SVM, SVM with RBF kernel | RF: 76.1; MLP: 74.5; XGB: 74.0; LightGBM: 73.1; qSOFA score: 56.0; RF (validation): 70.9 |
| Tsurumi A, 2023 | XGB, RF, Gaussian Naïve Bayes, SVM | Multibiomarker panel: 93.8 [95% CI 88.1–98.1]; Multibiomarker panel + TBSA: 96.5 [95% CI 93.9–99.9]; Multibiomarker panel + TBSA + inhalation injury status: 97.8 [95% CI 94.1–100.0] |
| Tzelves L, 2022 | Artificial NN | Bagging classifier: 87.4; Multinomial LR 76.8 |
| Viswanathan V, 2024 | RF, MLP, XGB, LightGBM | NA |
| Wang C-X, 2022 | Naive Bayes, RF, SVM, J48(C4.5), SMO | SVM: 89.0; XGB: 89.0; RF: 85.0 |
| Wang J, 2022 | LASSO, XGB, RF, Ensemble Model combining Algorithms | RF: 90.1; SVM: 92.9; SVM with RBF kernel: 93.6 |
| Wang Y, 2023 | J48 (C4.5), RF, Multinomial LR, k-NN, MLP | 81.1 [95% CI 73.1–89.1] |
| Wang Z, 2021 | Multinomial LR; bagging classifier | Discovery Population: 89.0 [95% CI 88.5–89.3]; Replication Population: 85.0 [95% CI 84.1–86.1] |
| Wong J-G, 2020 | RIPPER (JRip), RF, MLP, Classification via Regression, REPT | LASSO 70.0 [95% CI 62.0–77.0]; CART 67.0 [95% CI 59.0–74.0] |
| Yu J, 2023 | Risk-Calibrated Supersparse Linear Integer Model | S. aureus: training: 85.0, testing: 87.0; K. pneumoniae: training: 82.0, testing: 82.0 |
| Zhang Y-M, 2023 | NN | 91.0 |

AMP Ampicillin, AI Artificial Intelligence, CAR Carbapenems, CART Classification and Regression Trees, C. Clostridioides, CIP Ciprofloxacin, CRE Carbapenem-Resistant Enterobacteriaceae, CR Carbapenem-Resistant, DT Decision Tree, E. Escherichia, eICU eICU Collaborative Research Database, ESBL Extended-Spectrum Beta-Lactamase, GBM Gradient Boosting Machine, GEN Gentamicin, GPT Glutamate Pyruvate Transaminase, GRU Gated Recurrent Unit, GNB Gram-Negative Bacteria, K. Klebsiella, KSSHIBA Kernelized Sparse Semi-Supervised Inter-Battery Bayesian Analysis, LASSO Least Absolute Shrinkage and Selection Operator, LDA Linear Discriminant Analysis, LEV Levofloxacin, LR Logistic Regression, LSTM Long Short-Term Memory, MDRO Multidrug-Resistant Organism, MIMIC Medical Information Mart for Intensive Care, ML Machine Learning, MLP Multilayer Perceptron, MRSA Methicillin-Resistant Staphylococcus Aureus, NIT Nitrofurantoin, NN Neural Network, nHABP Non-Ventilator-associated Hospital-Acquired Bacterial Pneumonia, P. Pseudomonas, PIKE Peak Information Kernel, POLY Polymyxins, PR Polynomial Regression, QUI Quinolones, RBF Radial Basis Function, REPT Reduced Error Pruning Tree, RF Random Forest, RIPPER Repeated Incremental Pruning to Produce Error Reduction, SHAP SHapley Additive exPlanations, SMO Sequential Minimal Optimization, SMOTE Synthetic Minority Over-sampling Technique, SMX Sulfamethoxazole, SVM Support Vector Machine, TBSA Total Burn Surface Area, TMP Trimethoprim, TZP Piperacillin/tazobactam, UTI Urinary Tract Infection, VABP Ventilator-Associated Bacterial Pneumonia, vHABP Ventilator-associated Hospital-Acquired Bacterial Pneumonia, VRE Vancomycin-Resistant Enterococci, WBC White Blood Cells, XGB Extreme Gradient Boosting

effects model (FEM) revealed an ES of 83.40 (95% CI: 83.40–83.40, $p < 0.001$) based on a total of 1,075,058 participants, though significant heterogeneity was observed ($I^2 = 100\%$, $p < 0.001$). Applying a random effects model (REM), the ES decreased to 72.28 (95% CI: 70.42–74.14, $p < 0.001$), reflecting lower predictive performance when accounting for study variability. Publication bias was detected through visual inspection of the funnel plot and confirmed by Egger's regression test (intercept: -484.04 , $p = 0.020$). These results are presented in Fig. 3 (a: Forest plot, b: Funnel plot).

Regarding accuracy, we evaluated 8 studies comprising 48 ML models. The FEM showed an ES of 76.15 (95% CI: 76.14–76.16, $p < 0.001$), again with high heterogeneity ($I^2 = 100\%$) and a total sample of 95,626 patients. The REM yielded a comparable ES of 74.97 (95% CI: 73.35–76.58, $p < 0.001$). No evidence of publication bias was found in this case, as indicated by the funnel plot and Egger's test (intercept: 31.69 , $p = 0.368$). Results are illustrated in Fig. 4 (a: Forest plot, b: Funnel plot).

For sensitivity, 10 studies with 51 models were included. The FEM showed an ES of 72.68 (95% CI: 72.67–72.70, $p < 0.001$), alongside substantial heterogeneity ($I^2 = 100\%$, $p < 0.001$). The REM, however, indicated an improved ES of 76.89 (95% CI: 71.90–81.89, $p < 0.001$), based on 69,982 patients. No publication bias was detected, as confirmed by Egger's test (intercept: 117.04 , $p = 0.113$). Corresponding plots are shown in Fig. 5.

When assessing specificity, 13 studies encompassing 59 models were evaluated. The FEM provided an ES of 99.84 (95% CI: 99.84–99.84, $p < 0.001$), although heterogeneity remained high ($I^2 = 100\%$, $p < 0.001$). The REM showed a significantly lower ES of 73.77 (95% CI: 67.87–79.67, $p < 0.001$), based on a total of 94,113 participants. Visual analysis of the funnel plot and Egger's test (intercept: 401.75 , $p < 0.001$) confirmed potential publication bias. These findings are presented in Fig. 6.

For NPV, 6 studies with 12 models were included. The FEM reported an ES of 81.16 (95% CI: 81.14–81.18, $p < 0.001$), with high heterogeneity ($I^2 = 100\%$, $p < 0.001$), and a sample size of 21,338. The REM demonstrated a slightly lower ES of 79.92 (95% CI: 76.54–83.31, $p < 0.001$). No publication bias was identified (Egger's test: intercept 70.68 , $p = 0.301$). Results are shown in Fig. 7.

Lastly, for PPV, 5 studies with 11 models were analyzed. The FEM yielded an ES of 56.83 (95% CI: 56.81–56.85, $p < 0.001$), with high heterogeneity ($I^2 = 100\%$, $p < 0.001$) based on 10,976 participants. The REM showed a higher ES of 69.41 (95% CI: 60.19–78.63, $p < 0.001$). No publication bias was detected (Egger's test: intercept 298.55 , $p = 0.533$). These results are shown in Fig. 8.

Sensitivity analysis

Sensitivity analysis by study design was not conducted, as all included studies were cross-sectional. Instead, we performed sensitivity analysis based on the type of ML models used. For AUC, when restricted to studies using DT models (CART, RF, J48 [C4.5], DT), 11 studies and 36 ML models were analyzed, covering a total of 291,984 patients. The FEM demonstrated an ES of 79.44 (95% CI: 79.44–79.44, $p < 0.001$), although there was considerable heterogeneity ($I^2 = 100\%$, $p < 0.001$). The REM showed a lower ES of 68.18 (95% CI: 64.10–72.25, $p < 0.001$) (Fig. 9a and S2). Conversely, studies employing boosted models (XGB, GBM, CatBoost, Boosted LR, Boosted DT), comprising 9 studies and 18 models with a sample of 324,381 patients, showed an ES of 78.26 (95% CI: 78.26–78.26, $p < 0.001$) under the FEM. The REM showed an ES of 74.05 (95% CI: 68.02–80.09, $p < 0.001$) (Fig. 9b and S3).

For sensitivity, studies using decision tree-based ML models (5 studies, 18 models) with a sample of 23,155 patients, showed an ES of 78.46 (95% CI: 78.43–79.49, $p < 0.001$) in the FEM, with high heterogeneity ($I^2 = 99.99\%$, $p < 0.001$). The REM produced an ES of 74.97 (95% CI: 71.20–78.74, $p < 0.001$) (Fig. S4). In contrast, 5 studies using general neural networks (Artificial NN, NN, MLP), with 10 models and a sample size of 18,307 patients, showed an ES of 60.98 (95% CI: 60.95–61.00, $p < 0.001$) for the FEM, again with substantial heterogeneity ($I^2 = 99.99\%$, $p < 0.001$). The REM indicated an improved ES of 75.76 (95% CI: 61.25–90.27, $p < 0.001$) (Figure S5).

For specificity, 6 studies using DT models (RF, CART, DT, J48) were included, covering 20 models and a sample of 24,211 patients. The FEM showed an ES of 51.89 (95% CI: 51.86–51.91, $p < 0.001$), with substantial heterogeneity ($I^2 = 100\%$, $p < 0.001$). The REM showed a higher ES of 71.83 (95% CI: 59.13–84.53, $p < 0.001$) (Figure S6).

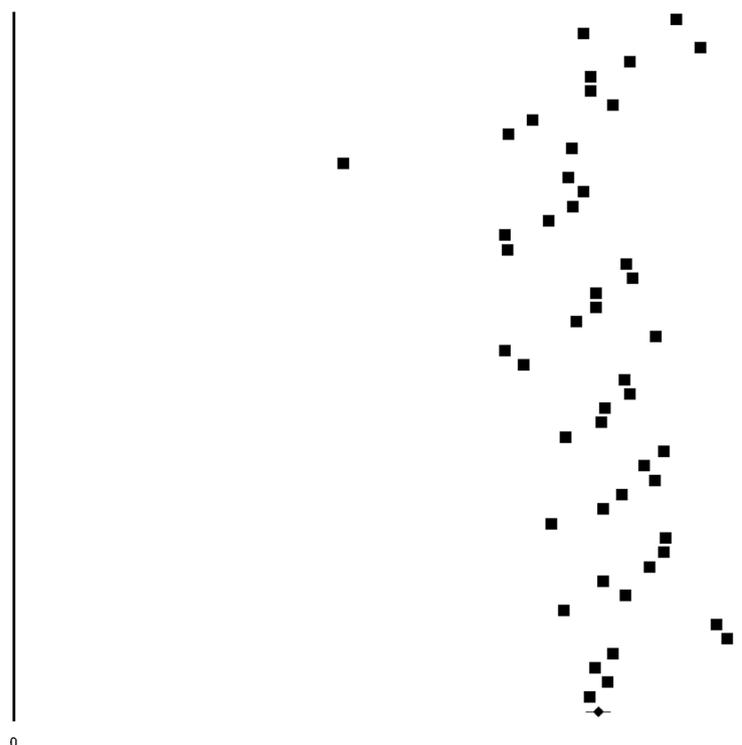
The high heterogeneity observed ($I^2 = 100\%$) can be attributed to differences in study designs, patient populations (e.g., ICU vs. general wards, adult vs. pediatric), and infection types (e.g., bloodstream vs. urinary infections). Additionally, variations in data sources (clinical vs. microbiological) contributed to the variability in outcomes.

Quality Assessment of included studies: QUADAS-AI

A detailed analysis of the risk of bias assessment and concerns regarding applicability was performed for each study and summarized in Figs. 10a and b. Risk of bias: 19 studies (24%) [13, 17, 26–28, 30, 32, 39–41, 53, 54, 56, 64, 67–70, 85] had a high risk of bias in patient selection mostly due to the lack of a clear rationale for its sample size and the unspecified data source. Most of the studies had a high risk of bias for index test (55%), which was most often due to the

a)

| | ES | 95% CI | Sig. | N |
|--|-------|--------------|-------|-------|
| Bolton W.-J, NN with SHAP 2024 | 85.00 | 84.96, 85.04 | 0.000 | 10362 |
| Bolton W.-J, Recurrent NN 2022 | 73.00 | 72.99, 73.01 | 0.000 | 18988 |
| Herman B, Artificial NN 2021 | 88.00 | 87.88, 88.12 | 0.000 | 644 |
| Herman B, DT 2021 | 79.00 | 78.85, 79.15 | 0.000 | 644 |
| Herman B, RF 2021 | 74.00 | 73.85, 74.15 | 0.000 | 644 |
| Herman B, XGB 2021 | 74.00 | 73.85, 74.15 | 0.000 | 644 |
| Hernández-Carnero A, LSTM1 2023 | 76.80 | 76.40, 77.20 | 0.000 | 755 |
| Hernández-Carnero A, LSTM2 2023 | 66.50 | 66.24, 66.76 | 0.000 | 643 |
| Hernández-Carnero A, LSTM3 2023 | 63.40 | 63.21, 63.59 | 0.000 | 749 |
| Hernández-Carnero A, LSTM4 2023 | 71.60 | 71.35, 71.85 | 0.000 | 749 |
| Hernández-Carnero A, LSTM5 2023 | 42.30 | 41.70, 42.90 | 0.000 | 483 |
| Hernández-Carnero A, LSTM6 2023 | 71.10 | 70.69, 71.51 | 0.000 | 708 |
| Hirano Y, XGB 2021 | 73.00 | 72.84, 73.16 | 0.000 | 809 |
| Mancini A, CatBoost 2020 | 71.70 | 71.54, 71.86 | 0.000 | 1486 |
| Mancini A, NN 2020 | 68.60 | 68.50, 68.70 | 0.000 | 1486 |
| Mancini A, SVM 2020 | 63.00 | 62.71, 63.29 | 0.000 | 1486 |
| Martinez-Aguero S, Bidirectional LSTM 2022 | 63.30 | 63.22, 63.38 | 0.000 | 3470 |
| Martinez-Aguero S, DT1 2019 | 78.60 | 78.50, 78.70 | 0.000 | 2177 |
| Martinez-Aguero S, DT2 2019 | 79.40 | 79.30, 79.50 | 0.000 | 1458 |
| Martinez-Aguero S, DT3 2019 | 74.60 | 74.49, 74.71 | 0.000 | 1582 |
| Martinez-Aguero S, DT4 2019 | 74.60 | 74.52, 74.68 | 0.000 | 2309 |
| Martinez-Aguero S, DT5 2019 | 72.10 | 71.53, 72.67 | 0.000 | 570 |
| Martinez-Aguero S, DT6 2019 | 82.30 | 82.23, 82.37 | 0.000 | 1952 |
| Martinez-Aguero S, GRU 2022 | 63.00 | 62.90, 63.10 | 0.000 | 3470 |
| Martinez-Aguero S, LSTM 2022 | 65.40 | 65.27, 65.53 | 0.000 | 3470 |
| Martinez-Aguero S, MLP1 2019 | 78.30 | 78.26, 78.34 | 0.000 | 2177 |
| Martinez-Aguero S, MLP2 2019 | 79.00 | 78.90, 79.10 | 0.000 | 1458 |
| Martinez-Aguero S, MLP3 2019 | 75.80 | 75.73, 75.87 | 0.000 | 1582 |
| Martinez-Aguero S, MLP4 2019 | 75.40 | 75.34, 75.46 | 0.000 | 2309 |
| Martinez-Aguero S, MLP5 2019 | 70.80 | 70.29, 71.31 | 0.000 | 570 |
| Martinez-Aguero S, MLP6 2019 | 83.40 | 83.31, 83.49 | 0.000 | 1952 |
| Martinez-Aguero S, RF1 2019 | 80.80 | 80.75, 80.85 | 0.000 | 2177 |
| Martinez-Aguero S, RF2 2019 | 82.20 | 82.11, 82.29 | 0.000 | 1458 |
| Martinez-Aguero S, RF3 2019 | 78.00 | 77.91, 78.09 | 0.000 | 1582 |
| Martinez-Aguero S, RF4 2019 | 75.60 | 75.55, 75.65 | 0.000 | 2309 |
| Martinez-Aguero S, RF5 2019 | 68.90 | 68.33, 69.47 | 0.000 | 570 |
| Martinez-Aguero S, RF6 2019 | 83.60 | 83.54, 83.66 | 0.000 | 1952 |
| Martinez-Aguero S, k-NN1 2019 | 83.30 | 83.22, 83.38 | 0.000 | 2177 |
| Martinez-Aguero S, k-NN2 2019 | 81.50 | 81.43, 81.57 | 0.000 | 1458 |
| Martinez-Aguero S, k-NN3 2019 | 75.60 | 75.52, 75.68 | 0.000 | 1582 |
| Martinez-Aguero S, k-NN4 2019 | 78.40 | 78.34, 78.46 | 0.000 | 2309 |
| Martinez-Aguero S, k-NN5 2019 | 70.50 | 69.93, 71.07 | 0.000 | 570 |
| Martinez-Aguero S, k-NN6 2019 | 90.10 | 90.04, 90.16 | 0.000 | 1952 |
| Shang J.-S, NN 2000 | 91.50 | 91.33, 91.67 | 0.000 | 504 |
| de Vries S, RF 2022 | 76.80 | 76.73, 76.87 | 0.000 | 810 |
| de Vries S, SVM 2022 | 74.50 | 74.43, 74.57 | 0.000 | 810 |
| de Vries S, XGB 2022 | 76.20 | 76.14, 76.26 | 0.000 | 810 |
| de Vries S, k-NN 2022 | 73.80 | 73.74, 73.86 | 0.000 | 810 |
| Overall (random-effects model) | 74.97 | 73.35, 76.58 | 0.000 | 95626 |



b)

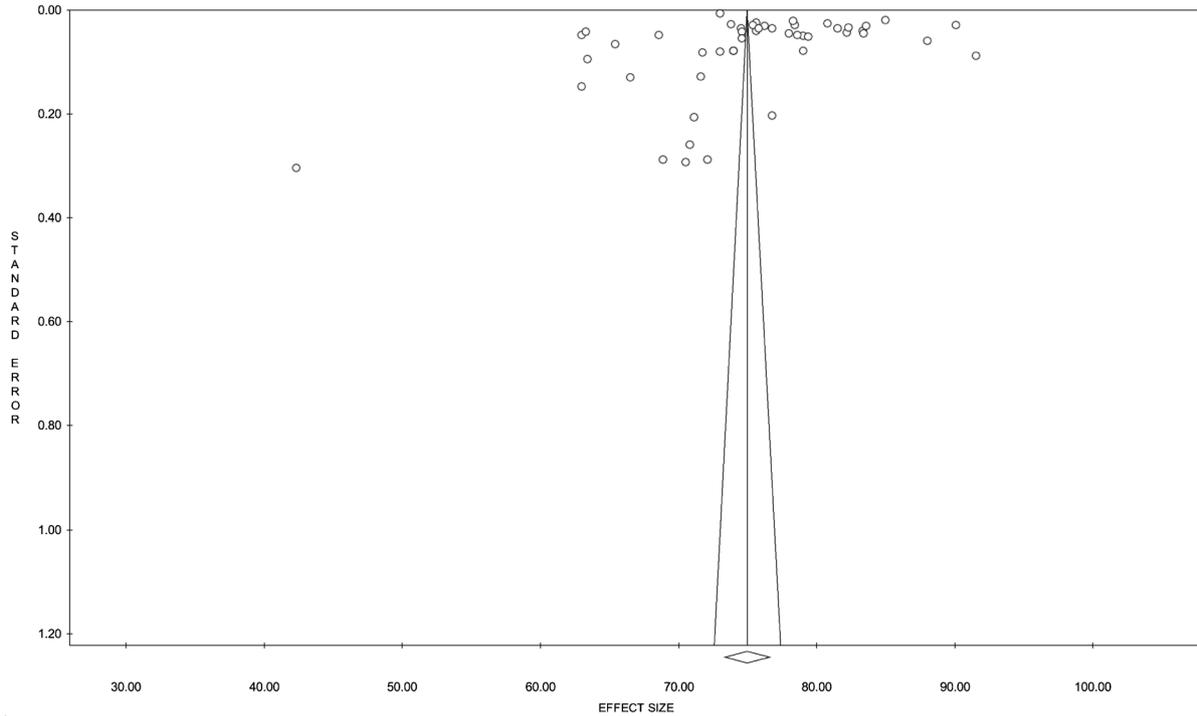
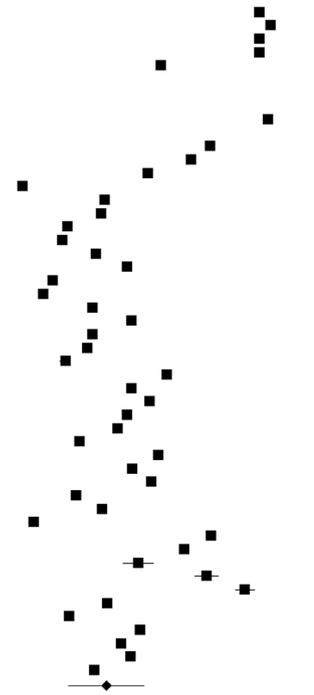


Fig. 4 a Forest plot and b funnel plot of the random effect model assessing the Accuracy

a)

| | ES | 95% CI | Stig. | N |
|--|-------|--------------|-------|-------|
| Ananda-Rajah M-R, J48(C4.5) 2017 | 96.90 | 96.42, 97.38 | 0.000 | 123 |
| Ananda-Rajah M-R, Naive Bayes 2017 | 98.40 | 98.03, 98.77 | 0.000 | 123 |
| Ananda-Rajah M-R, RF 2017 | 96.90 | 96.42, 97.38 | 0.000 | 123 |
| Ananda-Rajah M-R, SMO 2017 | 96.90 | 96.42, 97.38 | 0.000 | 123 |
| Herman B, Artificial NN 2021 | 84.00 | 83.75, 84.25 | 0.000 | 644 |
| Herman B, DT 2021 | 44.00 | 43.67, 44.33 | 0.000 | 644 |
| Herman B, RF 2021 | 25.00 | 24.71, 25.29 | 0.000 | 644 |
| Herman B, XGB 2021 | 25.00 | 24.71, 25.29 | 0.000 | 644 |
| Hirano Y, XGB 2021 | 98.00 | 97.93, 98.07 | 0.000 | 809 |
| Lee A-L-H, NN 2021 | 37.50 | 37.46, 37.54 | 0.000 | 5625 |
| Mancini A, CatBoost 2020 | 90.40 | 90.09, 90.71 | 0.000 | 1486 |
| Mancini A, NN 2020 | 88.00 | 87.61, 88.39 | 0.000 | 1486 |
| Mancini A, SVM 2020 | 82.30 | 82.09, 82.51 | 0.000 | 1486 |
| Martinez-Aguero S, Bidirectional LSTM 2022 | 65.90 | 65.77, 66.03 | 0.000 | 3470 |
| Martinez-Aguero S, DT1 2019 | 76.60 | 76.49, 76.71 | 0.000 | 2177 |
| Martinez-Aguero S, DT2 2019 | 76.20 | 76.04, 76.36 | 0.000 | 1458 |
| Martinez-Aguero S, DT3 2019 | 71.80 | 71.65, 71.95 | 0.000 | 1582 |
| Martinez-Aguero S, DT4 2019 | 71.10 | 70.96, 71.24 | 0.000 | 2309 |
| Martinez-Aguero S, DT5 2019 | 75.50 | 74.85, 76.15 | 0.000 | 570 |
| Martinez-Aguero S, DT6 2019 | 79.60 | 79.50, 79.70 | 0.000 | 1952 |
| Martinez-Aguero S, GRU 2022 | 69.90 | 69.71, 70.09 | 0.000 | 3470 |
| Martinez-Aguero S, LSTM 2022 | 68.60 | 68.39, 68.81 | 0.000 | 3470 |
| Martinez-Aguero S, MLP1 2019 | 75.00 | 74.95, 75.05 | 0.000 | 2177 |
| Martinez-Aguero S, MLP2 2019 | 80.20 | 80.09, 80.31 | 0.000 | 1458 |
| Martinez-Aguero S, MLP3 2019 | 75.10 | 74.97, 75.23 | 0.000 | 1582 |
| Martinez-Aguero S, MLP4 2019 | 74.40 | 74.32, 74.48 | 0.000 | 2309 |
| Martinez-Aguero S, MLP5 2019 | 71.50 | 70.72, 72.28 | 0.000 | 570 |
| Martinez-Aguero S, MLP6 2019 | 84.80 | 84.72, 84.88 | 0.000 | 1952 |
| Martinez-Aguero S, RF1 2019 | 80.20 | 80.12, 80.28 | 0.000 | 2177 |
| Martinez-Aguero S, RF2 2019 | 82.50 | 82.37, 82.63 | 0.000 | 1458 |
| Martinez-Aguero S, RF3 2019 | 79.60 | 79.44, 79.76 | 0.000 | 1582 |
| Martinez-Aguero S, RF4 2019 | 78.30 | 78.20, 78.40 | 0.000 | 2309 |
| Martinez-Aguero S, RF5 2019 | 73.40 | 72.86, 73.94 | 0.000 | 570 |
| Martinez-Aguero S, RF6 2019 | 83.60 | 83.51, 83.69 | 0.000 | 1952 |
| Martinez-Aguero S, k-NN1 2019 | 80.30 | 80.22, 80.38 | 0.000 | 2177 |
| Martinez-Aguero S, k-NN2 2019 | 82.70 | 82.59, 82.81 | 0.000 | 1458 |
| Martinez-Aguero S, k-NN3 2019 | 72.90 | 72.77, 73.03 | 0.000 | 1582 |
| Martinez-Aguero S, k-NN4 2019 | 76.30 | 76.19, 76.41 | 0.000 | 2309 |
| Martinez-Aguero S, k-NN5 2019 | 67.40 | 66.79, 68.01 | 0.000 | 570 |
| Martinez-Aguero S, k-NN6 2019 | 90.50 | 90.42, 90.58 | 0.000 | 1952 |
| Shang J-S, NN 2000 | 87.10 | 86.82, 87.38 | 0.000 | 504 |
| Tsunami A, LASSO1 2023 | 81.00 | 78.97, 83.03 | 0.000 | 82 |
| Tsunami A, LASSO2 2023 | 90.00 | 88.40, 91.60 | 0.000 | 82 |
| Tsunami A, LASSO3 2023 | 95.00 | 93.68, 96.32 | 0.000 | 82 |
| Wong J-G, CART 2020 | 77.00 | 76.70, 77.30 | 0.000 | 715 |
| Wong J-G, LASSO 2020 | 72.00 | 71.72, 72.28 | 0.000 | 715 |
| de Vries S, RF 2022 | 81.30 | 81.22, 81.38 | 0.000 | 810 |
| de Vries S, SVM 2022 | 78.80 | 78.71, 78.89 | 0.000 | 810 |
| de Vries S, XGB 2022 | 80.00 | 79.92, 80.08 | 0.000 | 810 |
| de Vries S, k-NN 2022 | 75.30 | 75.22, 75.38 | 0.000 | 810 |
| Overall (random-effects model) | 76.89 | 71.90, 81.89 | 0.000 | 69982 |

0



b)

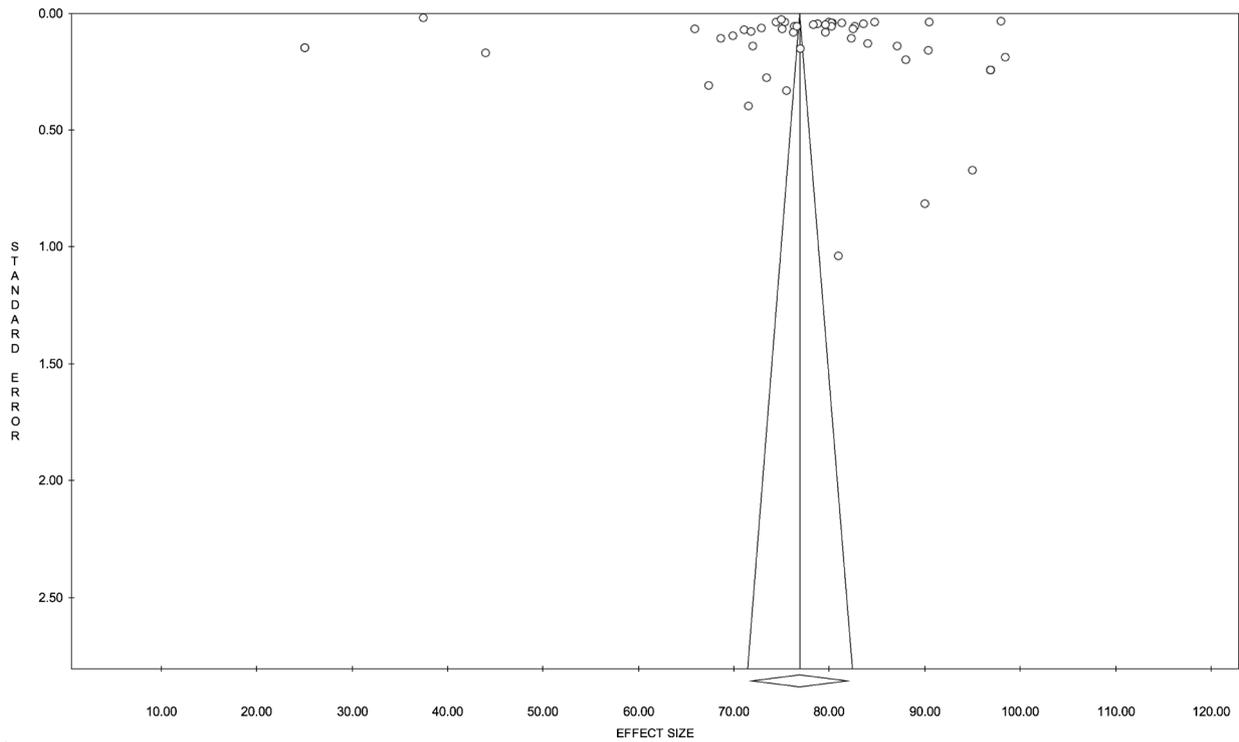
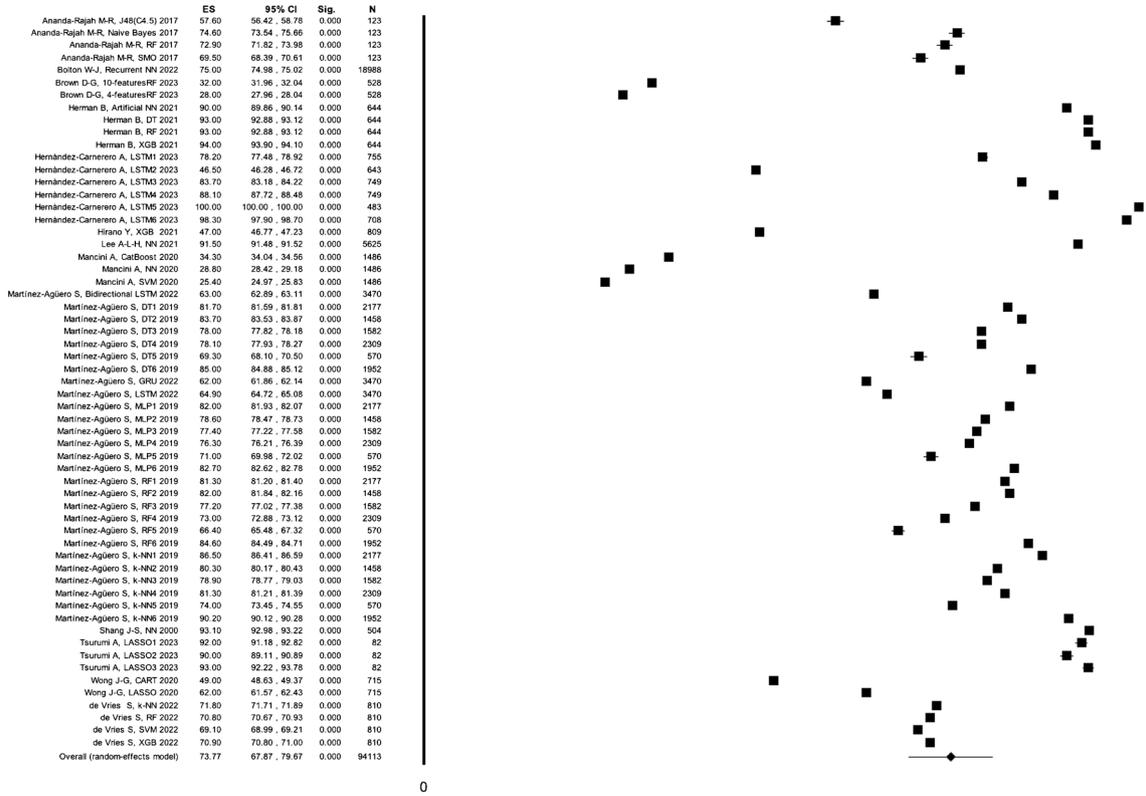


Fig. 5 a Forest plot and b funnel plot of the random effect model assessing the Sensitivity

a)



b)

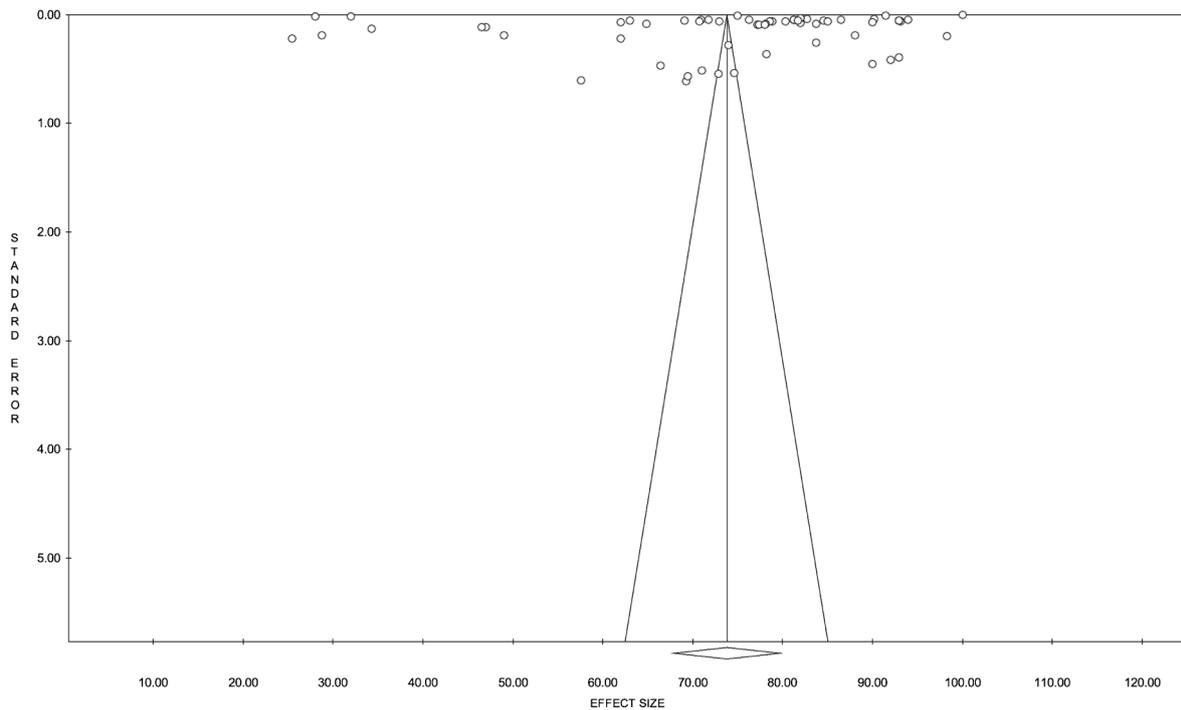
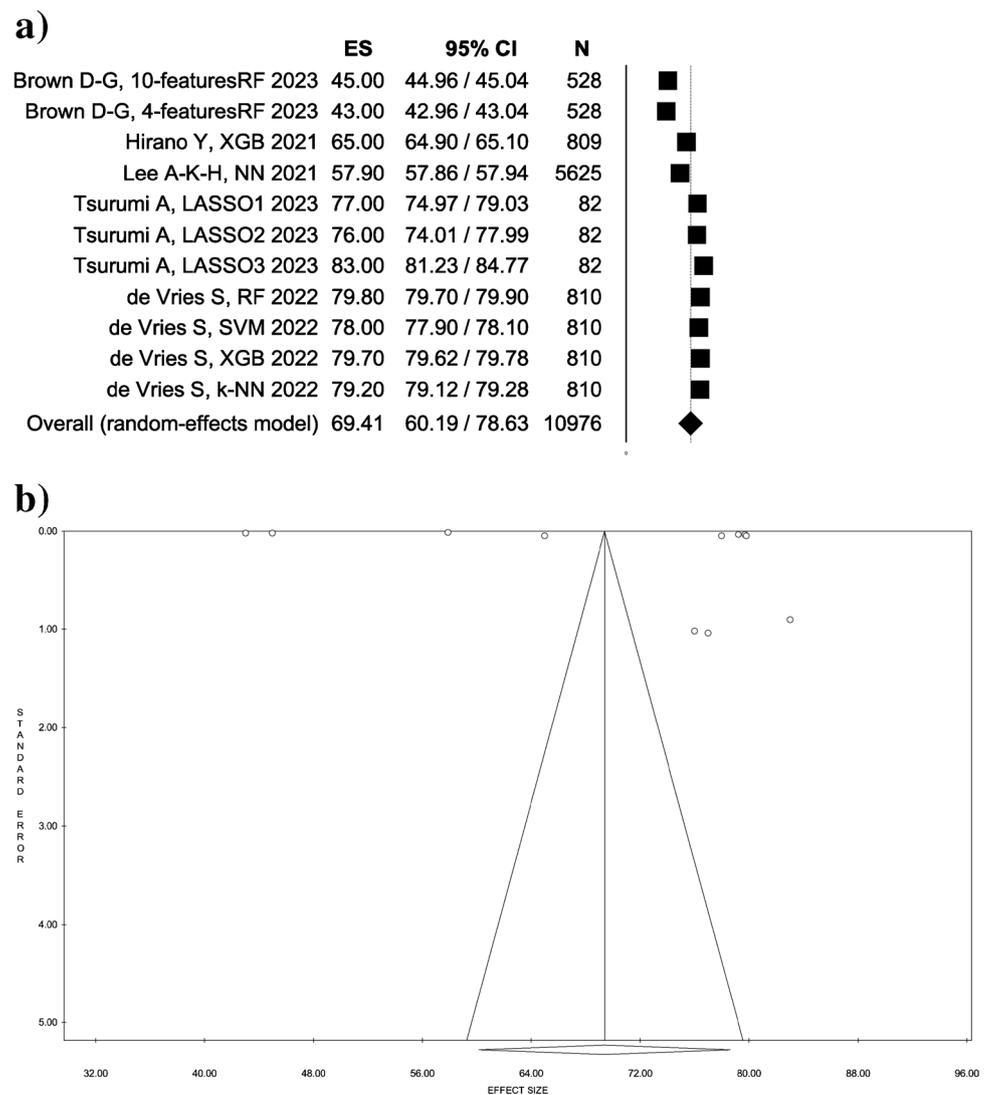


Fig. 6 a Forest plot and b funnel plot of the random effect model assessing the Specificity

Fig. 7 **a** Forest plot and **b** funnel plot of the random effect model assessing the PPV

lack of validation or testing on external data. In the reference standard domain, which assesses whether the method used as the gold standard for diagnosis or outcome measurement is reliable and applied appropriately, 10% of articles [26, 31, 39, 53, 68, 74, 75, 89] were at a high risk of bias. Finally, the risk of bias in the flow and timing domain was high in 18% of studies [12, 13, 17, 23, 32, 34, 38, 39, 53, 54, 66, 89, 90, 92]. Applicability concerns: In the patient selection domain, concerns about applicability were low only in 28% of the included studies [12–14, 16, 23, 31, 33, 34, 36, 37, 59, 63, 66, 67, 69, 71, 72, 85, 86, 90, 92, 93]. In the index test domain, concerns about applicability were high in 50% of the studies due to the lack of detail on the construct or architecture of the algorithm. Finally, in the reference standard domain, concerns about applicability were high in 14% of studies [26, 27, 31, 41, 53, 54, 75, 78, 80, 89, 94]. The main reasons for lower applicability included using surrogate or proxy measures instead of gold-standard diagnostic tests,

reliance on internal validation only and patient populations that may not be representative of the broader clinical setting. Additionally, issues like improper patient flow, where inclusion or exclusion criteria are unclear, or where the patient population does not mirror real-world clinical practice, could introduce bias and further reduce applicability.

Discussion

Interpretation of the results

The current systematic review of the literature assessed a total of more than 3 thousand studies, ultimately including 80 after the selection process. This data allows us to understand that, despite the topic being relatively new, the first study dates back to 2000, it is nonetheless gaining significant interest from the scientific community.

Fig. 8 **a** Forest plot and **b** funnel plot of the random effect model assessing the NPV

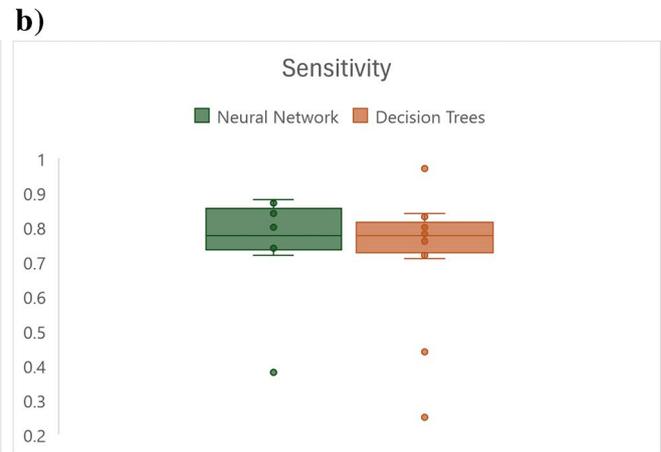
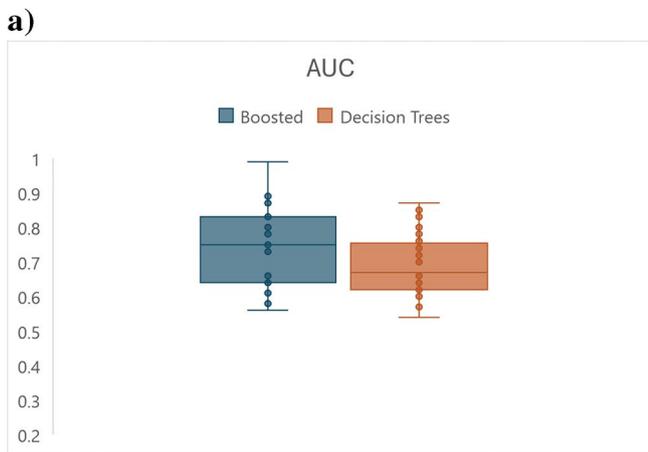
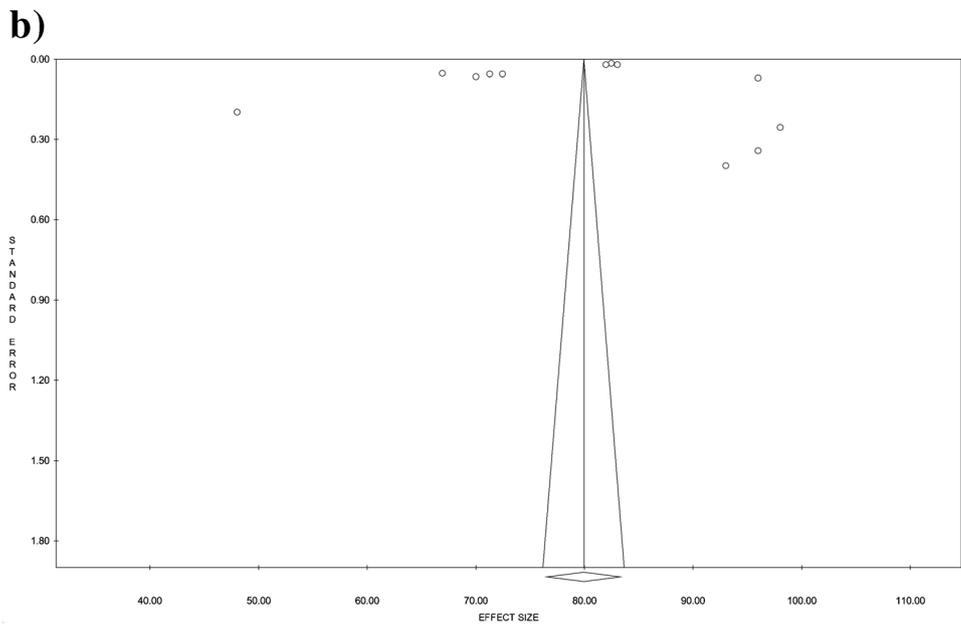
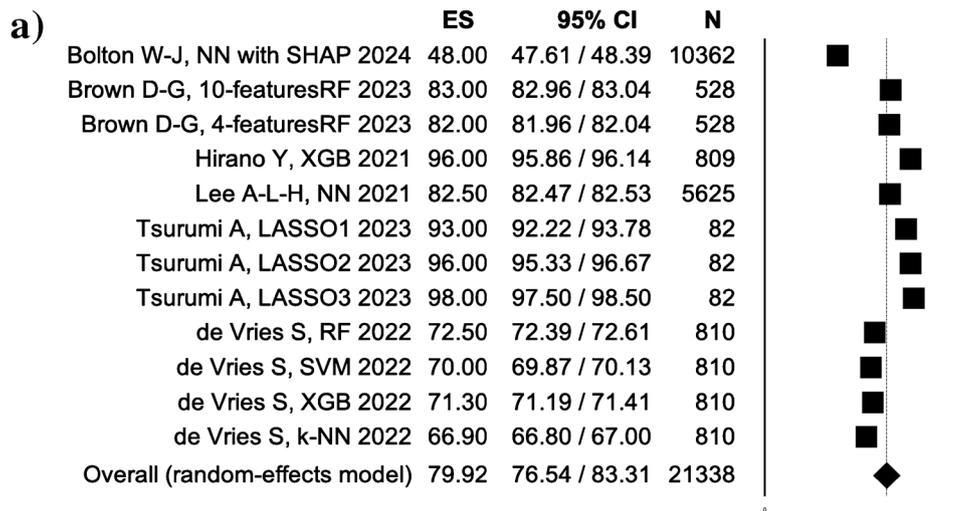
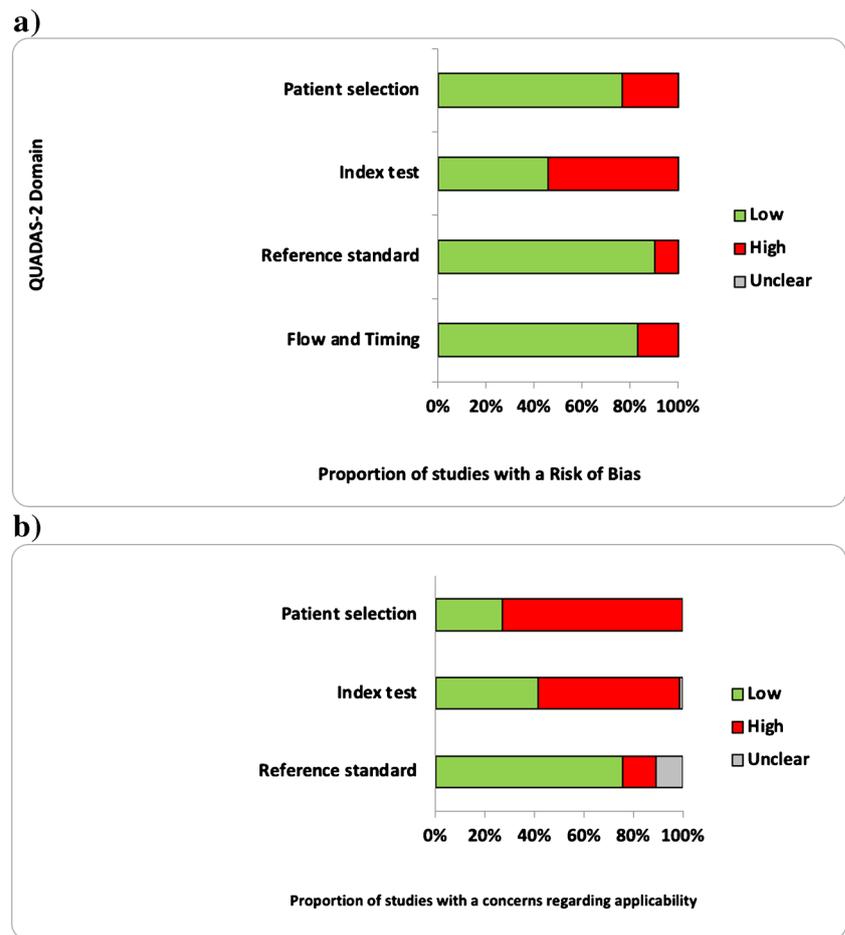


Fig. 9 **a** Sensitivity analysis for the area under the curve (AUC); and **b** for the sensitivity

Fig. 10 **a** Proportion of studies with a Risk of bias; and **b** with concerns regarding applicability



However, at the same time, the total number of studies that could be included in the meta-analytical evaluation was 29, highlighting the considerable heterogeneity in how data was collected and analyzed in each individual original study. This meta-analysis and systematic review demonstrated that ML models consistently show strong predictive performance in AMR prediction across different settings, as evidenced by the high AUC scores and diagnostic accuracy metrics. However, there was significant heterogeneity across the included studies, with variability in model performance depending on the algorithm used, the clinical setting, and the infection type. The REM consistently showed lower ES for AUC, accuracy, sensitivity, specificity, and predictive values, emphasizing the need to account for study variability in pooled analyses. The sensitivity analysis, which stratified results by the type of ML model employed, provided deeper insight into the differences between models.

The analysis showed that DT models, such as RF, CART, J48, and traditional DT, generally performed well, achieving consistent ES across metrics. For example, DT models showed a strong AUC in both FEMs (ES = 79.44) and REMs (ES = 68.18). DT models tend to perform well in clinical

applications because of their relative simplicity and interpretability [95]. In the context of AMR prediction, decision trees may outperform other models because their structure mirrors clinical decision-making processes, making them easy to validate and implement in real-world settings [96]. This could explain why DT models often exhibit high AUC and specificity in predicting AMR. One of the key determinants of model performance is the type and number of features included [97]. The studies included in this review had a wide range of feature numbers, from as few as five features to over 43,000. Models that can effectively select and prioritize important features, such as LASSO for dimensionality reduction or RF models that rank feature importance, are more likely to generalize well [98]. RF models, the most frequently used in the studies, inherently manage feature importance and handle large feature sets effectively. These capabilities contribute to their strong performance in predicting AMR across various datasets.

Boosted models similarly excel when feature selection is well-curated, as they iteratively refine the contribution of each feature to the prediction. Boosted models demonstrated superior performance compared to decision tree models in certain metrics, particularly AUC (ES = 74.05 under REMs),

indicating that these models might be better suited for AMR prediction in larger, more complex datasets. Despite their high predictive power, these models require vast amounts of high-quality data [99, 100]. Obtaining such data in healthcare is challenging due to privacy concerns, limited access to comprehensive datasets, and inconsistent record-keeping, particularly in underdeveloped regions [101–104]. Additionally, the temporal nature of medical data, such as evolving disease patterns, poses a challenge for models that rely on static data representations. While advanced architectures like bidirectional LSTMs attempt to address this, temporal dependencies in healthcare data remain complex [102]. These factors contribute to the mixed results seen in the application of neural networks despite their theoretical potential.

NN models, such as general ANNs, RNNs, and LSTM models, demonstrated inconsistent performance compared to decision tree and boosted models. Although neural networks are highly flexible and capable of handling large datasets, they often require substantial data preprocessing and hyperparameter tuning [105]. This might explain why, despite their theoretical potential, neural networks achieved lower ES values in the FEM for sensitivity (ES = 60.98) but showed an improved ES under REMs (ES = 75.76), particularly in studies that used more advanced recurrent architectures such as bidirectional LSTMs.

The predominance of internal validation suggests that some models may have been overfitted to specific datasets. Internal validation, while useful, can inflate performance metrics such as AUC or accuracy by tailoring the model too closely to the training data. In contrast, external validation provides a more rigorous test of model generalizability. This could explain why models that underwent external validation, such as those using decision trees and RF, showed lower but more reliable performance metrics in the REMs. The limited use of external validation in healthcare-related ML models points to a relevant challenge in evaluating true model performance.

The sensitivity analysis confirmed high heterogeneity ($I^2 = 100\%$ across nearly all models and metrics). This likely stems from differences in study design, patient populations, infection types, and the algorithms used. For example, studies focused on high-risk populations (e.g., ICU patients) or specific infections (e.g., genitourinary infections) may produce different results compared to those with more general patient populations. Additionally, low standardization in ML methods might also contribute to high heterogeneity. Publication bias, especially noted in specificity measures, suggests that studies reporting poor performance may have been underreported. This may skew the apparent superiority of certain models, like RF, in specific performance metrics, even if their generalizability to diverse clinical settings is less clear. This bias should be considered when interpreting

these findings. Overall, the results confirm that ML models, particularly decision trees and boosted models, perform well in predicting AMR, but their utility is influenced by the specific clinical context, model selection, and dataset characteristics.

Comparison with existing literature

The findings of this review are consistent with existing literature on the use of ML in healthcare, particularly in infectious disease management. Previous studies have shown the utility of ML algorithms, such as RF, SVM, and NN in improving the diagnostic capabilities of predictive models in a variety of clinical settings. Similar to prior reviews [7, 106], this study found that RF and boosted models, such as XGBoost, performed particularly well in terms of AUC, accuracy, and specificity. The widespread application of ML in AMR prediction aligns with earlier findings that ML can assist in stratifying patients and guiding therapeutic decisions in complex cases, such as bloodstream infections and respiratory infections. However, the heterogeneity observed in this review mirrors challenges noted in the broader literature, where differences in data quality, feature selection, and study designs lead to variable model performance. This underscores the need for standardized reporting and model validation practices to improve the comparability of ML models across studies.

Public health implications

This systematic review emphasizes the significant role ML models could play in AMS programs to combat AMR, offering faster, accurate predictions that optimize antibiotic use and minimize the spread of resistance. Integrating ML into clinical settings can reduce reliance on traditional culture tests, which take days to yield results, allowing clinicians to make informed decisions within hours and avoid premature antibiotic use. This aligns with the One Health approach, as reducing AMR in humans also benefits animal and environmental health. The review shows ML's potential in precision medicine and infection control, especially in high-risk environments like ICUs, with 65% of studies focusing on hospital settings. Additionally, ML can help identify facility-level antibiotic usage predictors, aiding in targeted infection control measures. However, due to publication bias in some studies, further independent validation of these ML models is necessary before broader implementation.

Future directions

Future research should focus on addressing the limitations identified in this review, including the need for more external validation of ML models. Future studies should

aim to validate ML models in diverse clinical settings, particularly in ambulatory and emergency care environments, which were underrepresented in the current review. Additionally, there is a need for more research focused on pediatric populations, as only 11.3% of the studies included children. Another priority for future research should be the development of models that integrate data from multiple institutions, enhancing their generalizability. Finally, the detection of publication bias in some areas suggests the need for more transparency in reporting, and future research should prioritize open data sharing and the use of standardized metrics for evaluating ML model performance.

Strengths and limitations

The main strength of this review is its comprehensive analysis of ML models for AMR prediction across diverse regions and settings, covering over 1.3 million patients to enhance generalizability. The systematic review with meta-analysis rigorously evaluates AUC, accuracy, sensitivity, specificity, and predictive values, highlighting the strengths and weaknesses of different models. Conducted per international guidelines, this work ensures rigor and transparency. However, high study heterogeneity and publication bias, especially in specificity, limit the ability to draw definitive conclusions.

Conclusion

This systematic review and meta-analysis demonstrate the significant potential of ML models to enhance AMS by improving diagnostic accuracy and predicting AMR. ML models, particularly DT, RF, and boosted algorithms, showed strong predictive performance and could serve as valuable tools in assisting AMS teams, and helping to control the global rise of AMR. However, several limitations emerged from this review. The generalizability of these models remains a concern, as many studies lacked standardization in methodologies and validation approaches, potentially limiting their applicability in diverse clinical settings. Additionally, the complexity and interpretability of some ML models pose challenges to their adoption in practice, as they may not be easily understood or trusted by clinicians. In conclusion, while ML models hold great promise for improving AMS, further research is needed to ensure their effective implementation. Standardized guidelines should be introduced to ensure consistency across future studies, and these models should be tested in randomized controlled trials or real-world settings to validate their practical utility in diverse healthcare environments.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s10096-024-05027-y>.

Author contributions F. P. and V. G. contributed to the study conception and design. Material preparation, the literature search and data analysis were performed by all the authors. The first draft of the manuscript was written by all the authors. F. P. and V. G. critically revised the work. All authors read and approved the final manuscript.

Funding The authors declare that no funds, grants, or other support were received during the preparation of this manuscript.

Data availability Data are available from the corresponding author upon reasonable request (risk of bias assessment data).

Declarations

Competing interests The authors declare no competing interests.

References

- Murray CJ, Ikuta KS, Sharara F, Swetschinski L, Robles Aguilar G, Gray A et al (2022) Global burden of bacterial antimicrobial resistance in 2019: a systematic analysis. *Lancet* 399:629–655
- Zhang C, Fu X, Liu Y, Zhao H, Wang G (2024) Burden of infectious diseases and bacterial antimicrobial resistance in China: a systematic analysis for the global burden of disease study 2019. *Lancet Reg Health West Pac* 43:1–12
- Majumder MAA, Rahman S, Cohall D, Bharatha A, Singh K, Haque M et al (2020) Antimicrobial stewardship: Fighting antimicrobial resistance and protecting global public health. *Infect Drug Resist* 13:4713–4738
- Aiesh BM, Nazzal MA, Abdelhaq AI, Abutaha SA, Zyoud SH, Sabateen A (2023) Impact of an antibiotic stewardship program on antibiotic utilization, bacterial susceptibilities, and cost of antibiotics. *Sci Rep.* 13:1–9. <https://doi.org/10.1038/s41598-023-32329-6>
- Fanelli U, Chiné V, Pappalardo M, Gismondi P, Esposito S (2020) Improving the quality of hospital antibiotic use: impact on multidrug-resistant bacterial infections in children. *Front Pharmacol* 11:2019–2021
- Ali T, Ahmed S, Aslam M (2023) Artificial intelligence for antimicrobial resistance prediction: challenges and opportunities towards practical implementation. *Antibiotics* 12:523
- Pinto-de-Sá R, Sousa-Pinto B, Costa-de-Oliveira S (2024) Brave new world of artificial intelligence: its use in antimicrobial stewardship—a systematic review. *Antibiotics* 13:307
- Guni A, Sounderajah V, Whiting P, Bossuyt P, Darzi A, Ashrafian H (2024) A revised tool for the quality assessment of diagnostic accuracy studies utilising AI: protocol for QUADAS-AI. *JMIR Res Protoc* 13:e58202
- Yarahuan JKW, Kisvarday S, Kim E, Yan AP, Nakamura MM, Jones SB et al (2024) An Algorithm to Assess Guideline Concordance of Antibiotic Choice in Community-Acquired Pneumonia. *Hosp Pediatr* 14:137–145
- Agbaria AH, Salman A, Beck G, Lapidot I, Rich DH, Kapelushnik J et al (2019) Potential of bacterial infection diagnosis using infrared spectroscopy of WBC and machine learning algorithms. *Optics InfoBase Conference Papers. Part F142-:2024*
- Imamović E, Deumić A, Khouly A, Pisl KT, Avdić E, Hukić M et al (2021) Prediction of multi-drug resistance in *Escherichia coli* using machine learning algorithms. *IFMBE Proc* 84:155–163
- de Vries S, ten Doesschate T, Totté JEE, Heutz JW, Loeffen YGT, Oosterheert JJ et al (2022) A semi-supervised decision support

- system to facilitate antibiotic stewardship for urinary tract infections. *Comput Biol Med* 146:105621
13. Shi ZY, Hon JS, Cheng CY, Chiang HT, Huang HM (2022) Applying machine learning techniques to the audit of antimicrobial prophylaxis. *Appl Sci (Switzerland)* 12:2586
 14. Moehring RW, Phelan M, Lofgren E, Nelson A, Dodds Ashley E, Anderson DJ et al (2021) Development of a machine learning model using electronic health record data to identify antibiotic use among hospitalized patients. *JAMA Netw Open* 4:1–12
 15. Beaudoin M, Kabanza F, Nault V, Valiquette L (2016) Evaluation of a machine learning capability for a clinical decision support system to enhance antimicrobial stewardship programs. *Artif Intell Med* 68:29–36. <https://doi.org/10.1016/j.artmed.2016.02.001>
 16. Feretzakis G, Sakagianni A, Loupelis E, Kalles D, Skarmoutsou N, Martsoukou M et al (2021) Machine learning for antibiotic resistance prediction: A prototype using off-the-shelf techniques and entry-level data to guide empiric antimicrobial therapy. *Healthc Inform Res* 27:214–221
 17. Bystritsky RJ, Beltran A, Young AT, Wong A, Hu X, Doernberg SB (2020) Machine learning for the prediction of antimicrobial stewardship intervention in hospitalized patients receiving broad-spectrum agents. *Infect Control Hosp Epidemiol* 41:1022–1027
 18. Mancini A, Vito L, Marcelli E, Piangerelli M, De Leone R, Pucciarelli S et al (2020) Machine learning models predicting multidrug resistant urinary tract infections using “dsaaS.” *BMC Bioinforma* 21:1–12. <https://doi.org/10.1186/s12859-020-03566-7>
 19. Stracy M, Snitser O, Yelin I, Amer Y, Parizade M, Katz R et al (1979) Minimizing treatment-induced emergence of antibiotic resistance in bacterial infections. *Science* 202(375):889–894
 20. Corbin CK, Sung L, Chattopadhyay A, Noshad M, Chang A, Deresinski S et al (2022) Personalized antibiograms for machine learning driven antibiotic selection. *Commun Med* 2:38
 21. Eickelberg G, Sanchez-Pinto LN, Luo Y (2020) Predictive modeling of bacterial infections and antibiotic therapy needs in critically ill adults. *J Biomed Inform* 109:103540. <https://doi.org/10.1016/j.jbi.2020.103540>
 22. Goodman KE, Heil EL, Claeyss KC, Banoub M, Bork JT (2022) Real-world antimicrobial stewardship experience in a large academic medical center: using statistical and machine learning approaches to identify intervention “Hotspots” in an antibiotic audit and feedback program. *Open Forum Infect Dis* 9:ofac289
 23. Tzelves L, Lazarou L, Feretzakis G, Kalles D, Mourmouris P, Loupelis E et al (2022) Using machine learning techniques to predict antimicrobial resistance in stone disease patients. *World J Urol* 40:1731–6. <https://doi.org/10.1007/s00345-022-04043-x>
 24. Oonsivilai M, Mo Y, Luangasanatip N, Lubell Y, Miliya T, Tan P et al (2018) Using machine learning to guide targeted and locally-tailored empiric antibiotic prescribing in a children’s hospital in Cambodia. *Wellcome Open Res* 3:1–18
 25. Bolton WJ, Rawson TM, Hernandez B, Wilson R, Antcliffe D, Georgiou P et al (2022) Machine learning and synthetic outcome estimation for individualised antimicrobial cessation. *Front Digit Health* 4:1–12
 26. Wong JG, Aung AH, Lian W, Lye DC, Ooi CK, Chow A (2020) Risk prediction models to guide antibiotic prescribing: a study on adult patients with uncomplicated upper respiratory tract infections in an emergency department. *Antimicrob Resist Infect Control* 9:1–11
 27. Viswanathan V, Govindan S, Selvaraj B, Rupert S, Kumar R (2024) A clinical study to evaluate autofluorescence imaging of diabetic foot ulcers using a novel artificial intelligence enabled noninvasive device. *Int J Low Extrem Wounds* 23:169–176
 28. Çağlayan Ç, Barnes SL, Pineles LL, Harris AD, Klein EY (2022) A data-driven framework for identifying intensive care unit admissions colonized with multidrug-resistant organisms. *Front Public Health* 10:1–17
 29. Wang Y, Wang G, Zhao Y, Wang C, Chen C, Ding Y et al (2023) A deep learning model for predicting multidrug-resistant organism infection in critically ill patients. *J Intensive Care* 11:1–11. <https://doi.org/10.1186/s40560-023-00695-y>
 30. McGuire RJ, Yu SC, Payne PRO, Lai AM, Vazquez-Guillamet MC, Kollef MH et al (2021) A pragmatic machine learning model to predict carbapenem resistance. *Antimicrob Agents Chemother* 65:1–10
 31. Tsurumi A, Flaherty PJ, Que YA, Ryan CM, Banerjee A, Chakraborty A et al (2023) A preventive tool for predicting bloodstream infections in children with burns. *Shock* 59:393–399
 32. Chan ALF, Chen JX, Wang HY (2006) Application of data mining to predict the dosage of vancomycin as an outcome variable in a teaching hospital population. *Int J Clin Pharmacol Ther* 44:533–8
 33. Cai T, Anceschi U, Prata F, Collini L, Brugnolli A, Migno S et al (2023) Artificial intelligence can guide antibiotic choice in recurrent UTIs and become an important aid to improve antimicrobial stewardship. *Antibiotics* 12:375
 34. Guerrero-López A, Sevilla-Salcedo C, Candela A, Hernández-García M, Cercenado E, Olmos PM et al (2023) Automatic antibiotic resistance prediction in *Klebsiella pneumoniae* based on MALDI-TOF mass spectra. *Eng Appl Artif Intell* 118:105644. <https://doi.org/10.1016/j.engappai.2022.105644>
 35. Abu-Aqil G, Sharaha U, Suleiman M, Riesenberger K, Lapidot I, Salman A et al (2022) Culture-independent susceptibility determination of *E. coli* isolated directly from patients’ urine using FTIR and machine-learning. *Analyst* 147:4815–23
 36. Nigo M, Rasmy L, Mao B, Kannadath BS, Xie Z, Zhi D (2024) Deep learning model for personalized prediction of positive MRSA culture using time-series electronic health records. *Nat Commun* 15:1–11
 37. Lee ALH, To CCK, Lee ALS, Chan RCK, Wong JSH, Wong CW et al (2021) Deep learning model for prediction of extended-spectrum beta-lactamase (ESBL) production in community-onset Enterobacteriaceae bacteraemia from a high ESBL prevalence multi-centre cohort. *Eur J Clin Microbiol Infect Dis* 40:1049–1061
 38. Schinkel M, Boerman AW, Paranjape K, Wiersinga WJ, Nanayakkara PWB (2023) Detecting changes in the performance of a clinical machine learning tool over time. *EBioMedicine* 97:104823. <https://doi.org/10.1016/j.ebiom.2023.104823>
 39. Huang TS, Lee SSI, Lee CC, Chang FC (2020) Detection of carbapenem-resistant *Klebsiella pneumoniae* on the basis of matrix-assisted laser desorption ionization time-of-flight mass spectrometry by using supervised machine learning approach. *PLoS One* 15:1–13
 40. Hassan N, Slight R, Weiand D, Morgan G, Vellinga A, Mamdough H et al (2023) Developing an AI-predictive model for predicting the likelihood of post-operative infection in surgical patients. *Int J Pharm Pract* 31:ii22-3
 41. Seheult JN, Stram MN, Contis L, Pontzer RE, Hardy S, Wertz W et al (2023) Development, evaluation, and multisite deployment of a machine learning decision tree algorithm to optimize urinalysis parameters for predicting urine culture positivity. *J Clin Microbiol* 61:e0029123

42. Herman B, Sirichokchatchawan W, Pongpanich S, Nantasenam C (2021) Development and performance of CUHASROBUST application for pulmonary rifampicin-resistance tuberculosis screening in Indonesia. *PLoS One*. 16:1–19. <https://doi.org/10.1371/journal.pone.0249243>
43. Bhavani S, Lonjers Z, Carey K, Afshar M, Gilbert E, Shah N et al (2024) The development and validation of a machine learning model to predict bacteremia and fungemia in hospitalized patients using electronic health record data. *Crit Care Med* 48:1–16
44. Rich SN, Jun I, Bian J, Boucher C, Cherabuddi K, Morris JG et al (2022) Development of a prediction model for antibiotic-resistant urinary tract infections using integrated electronic health records from multiple clinics in North-Central Florida. *Infect Dis Ther*. 11:1869–82. <https://doi.org/10.1007/s40121-022-00677-x>
45. Brown DG, Worby CJ, Pender MA, Brintz BJ, Ryan ET, Sridhar S et al (2023) Development of a prediction model for the acquisition of extended spectrum beta-lactam-resistant organisms in U.S. international travellers. *J Travel Med* 30:1–9
46. Liu Q, Chen Y, Xie P, Luo Y, Wang B, Meng Y et al (2024) Development of a predictive machine learning model for pathogen profiles in patients with secondary immunodeficiency. *BMC Med Inform Decis Mak*. 24:1–11. <https://doi.org/10.1186/s12911-024-02447-w>
47. Tran-The T, Heo E, Lim S, Suh Y, Heo KN, Lee EE et al (2024) Development of machine learning algorithms for scaling-up antibiotic stewardship. *Int J Med Inform*. 181:105300. <https://doi.org/10.1016/j.ijmedinf.2023.105300>
48. Takács AT, Bukva M, Bereczki C, Burián K, Terhes G (2023) Diagnosis of Epstein-Barr and cytomegalovirus infections using decision trees: an effective way to avoid antibiotic overuse in paediatric tonsillopharyngitis. *BMC Pediatr* 23:1–9
49. Hernández-Carnerero Á, Sánchez-Marrè M, Mora-Jiménez I, Soguero-Ruiz C, Martínez-Agüero S, Álvarez-Rodríguez J (2023) Dimensionality reduction and ensemble of LSTMs for antimicrobial resistance prediction. *Artif Intell Med*. 138:102508. <https://doi.org/10.1016/j.artmed.2023.102508>
50. Kong PH, Chiang CH, Lin TC, Kuo SC, Li CF, Hsiung CA et al (2022) Discrimination of Methicillin-Resistant *Staphylococcus aureus* by MALDI-TOF mass spectrometry with machine learning techniques in patients with *Staphylococcus aureus* Bacteremia. *Pathogens* 11:586
51. Liang Q, Zhao Q, Xu X, Zhou Y, Huang M (2022) Early prediction of carbapenem-resistant Gram-negative bacterial carriage in intensive care units using machine learning. *J Glob Antimicrob Resist*. 29:225–31. <https://doi.org/10.1016/j.jgar.2022.03.019>
52. Tacconelli E, Górská A, De Angelis G, Lammens C, Restuccia G, Schrenzel J et al (2020) Estimating the association between antibiotic exposure and colonization with extended-spectrum β -lactamase-producing Gram-negative bacteria using machine learning methods: a multicentre, prospective cohort study. *Clin Microbiol Infect* 26:87–94
53. Abu-Aqil G, Lapidot I, Salman A, Huleihel M (2023) Quick detection of proteus and pseudomonas in patients' urine and assessing their antibiotic susceptibility using infrared spectroscopy and machine learning. *Sensors*. 23:8132
54. Jiménez F, Palma J, Sánchez G, Marín D, Francisco Palacios MD, Lucía López MD (2020) Feature selection based multivariate time series forecasting: An application to antibiotic resistance outbreaks prediction. *Artif Intell Med*. 104:101818. <https://doi.org/10.1016/j.artmed.2020.101818>
55. Cavallaro M, Moran E, Collyer B, McCarthy ND, Green C, Keeling MJ (2023) Informing antimicrobial stewardship with explainable AI. *PLOS Digital Health*. 2:1–20. <https://doi.org/10.1371/journal.pdig.0000162>
56. Abu-Aqil G, Suleiman M, Lapidot I, Huleihel M, Salman A (2024) Infrared spectroscopy-based machine learning algorithms for rapid detection of *Klebsiella pneumoniae* isolated directly from patients' urine and determining its susceptibility to antibiotics. *Spectrochim Acta A Mol Biomol Spectrosc*. 314:124141. <https://doi.org/10.1016/j.saa.2024.124141>
57. Martínez-Agüero S, Soguero-Ruiz C, Alonso-Moral JM, Mora-Jiménez I, Álvarez-Rodríguez J, Marques AG (2022) Interpretable clinical time-series modeling with intelligent feature selection for early prediction of antimicrobial multidrug resistance. *Futur Gener Comput Syst* 133:68–83. <https://doi.org/10.1016/j.future.2022.02.021>
58. Ayyıldız H, Tuncer SA (2021) Is it possible to determine antibiotic resistance of *E. coli* by analyzing laboratory data with machine learning? *Turk J Biochem* 46:623–30
59. Wang Z, Wang HY, Chung CR, Horng JT, Lu JJ, Lee TY (2021) Large-scale mass spectrometry data combined with demographics analysis rapidly predicts methicillin resistance in *Staphylococcus aureus*. *Brief Bioinform* 22:1–12
60. Wang C, Wang Z, Wang HY, Chung CR, Horng JT, Lu JJ et al (2022) Large-scale samples based rapid detection of ciprofloxacin resistance in *Klebsiella pneumoniae* using machine learning methods. *Front Microbiol* 13:1–14
61. Hirano Y, Shinmoto K, Okada Y, Suga K, Bombard J, Murahata S et al (2021) Machine learning approach to predict positive screening of methicillin-resistant *staphylococcus aureus* during mechanical ventilation using synthetic dataset from MIMIC-IV database. *Front Med (Lausanne)* 8:1–9
62. Brokowski TJ, Chiang JN (2022) Machine learning for antibiotic stewardship in the treatment of *staphylococcus* bacterial infections. Available from: <https://www.embase.com/search/results?subaction=viewrecord&id=L2021629655&from=export>. <https://doi.org/10.1101/2022.11.28.22282797>. Accessed 30 Oct 2024
63. McFadden BR, Inglis TJJ, Reynolds M (2023) Machine learning pipeline for blood culture outcome prediction using Sysmex XN-2000 blood sample results in Western Australia. *BMC Infect Dis* 23:1–10. <https://doi.org/10.1186/s12879-023-08535-y>
64. Martínez-Agüero S, Mora-Jiménez I, Lérica-García J, Álvarez-Rodríguez J, Soguero-Ruiz C (2019) Machine learning techniques to identify antimicrobial resistance in the intensive care unit. *Entropy* 21:1–24
65. Garcia-Vidal C, Puerta-Alcalde P, Cardozo C, Orellana MA, Besanson G, Lagunas J et al (2021) Machine learning to assess the risk of multidrug-resistant gram-negative bacilli infections in febrile neutropenic hematological patients. *Infect Dis Ther* 10:971–983
66. Sophonsri A, Lou M, Ny P, Minejima E, Nieberg P, Wong-Beringer A (2023) Machine learning to identify risk factors associated with the development of ventilated hospital-acquired pneumonia and mortality: implications for antibiotic therapy selection. *Front Med (Lausanne)* 10:1–11
67. Rhodes NJ, Rohani R, Yarnold PR, Pawlowski AE, Malczynski M, Qi C et al (2023) Machine learning to stratify methicillin-resistant *staphylococcus aureus* risk among hospitalized patients with community-acquired pneumonia. *Antimicrob Agents Chemother* 67:1–10

68. Ravkin HD, Ravkin RM, Rubin E, Neshler L (2024) Machine-learning-based risk assessment tool to rule out empirical use of ESBL-targeted therapy in endemic areas. *J Hosp Infect.* 149:90–7. <https://doi.org/10.1016/j.jhin.2024.04.005>
69. Lapp Z, Wiens J (2021) Patient and microbial genomic factors associated with. *Am Soc Microbiol* 6:1–12
70. Jeon K, Kim JM, Rho K, Jung SH, Park HS, Kim JS (2022) Performance of a machine learning-based methicillin resistance of staphylococcus aureus identification system using MALDI-TOF MS and comparison of the accuracy according to SCCmec types. *Microorganisms* 10:1903
71. Bolton WJ, Wilson R, Gilchrist M, Georgiou P, Holmes A, Rawson TM (2024) Personalising intravenous to oral antibiotic switch decision making through fair interpretable machine learning. *Nat Commun* 15:1–13
72. Corbin CK, Medford RJ, Osei K, Chen JH (2020) Personalized Antibigrams: Machine Learning for Precision Selection of Empiric Antibiotics. *AMIA Jt Summits Transl Sci Proc.* 2020:108–15. Available from: <https://www.ncbi.nlm.nih.gov/pubmed/32477629%0A>, <https://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC7233062>. Accessed 30 Oct 2024
73. Lewin-Epstein O, Baruch S, Hadany L, Stein GY, Obolski U (2021) Predicting antibiotic resistance in hospitalized patients by applying machine learning to electronic medical records. *Clin Infect Dis* 72:E848–E855
74. Pascual-Sanchez L, Mora-Jimenez I, Martinez-Aguero S, Alvarez-Rodriguez J, Soguero-Ruiz C (2021) Predicting Multidrug Resistance Using Temporal Clinical Data and Machine Learning Methods. *Proceedings - 2021 IEEE International Conference on Bioinformatics and Biomedicine, BIBM 2021.* 2826–33
75. Al-khlifeh EM, Hassanat AB (2024) Predicting the distribution patterns of antibiotic-resistant microorganisms in the context of Jordanian cases using machine learning techniques. *J Appl Pharm Sci* 14:174–183
76. İlhanlı N, Park SY, Kim J, Ryu JA, Yardımcı A, Yoon D (2024) Prediction of antibiotic resistance in patients with a urinary tract infection: algorithm development and validation. *JMIR Med Inform* 12:1–11
77. Liang Q, Ding S, Chen J, Chen X, Xu Y, Xu Z et al (2024) Prediction of carbapenem-resistant gram-negative bacterial bloodstream infection in intensive care unit based on machine learning. *BMC Med Inform Decis Mak.* 24:1–11. <https://doi.org/10.1186/s12911-024-02504-4>
78. Yu J, Lin HH, Tseng KH, Lin YT, Chen WC, Tien N et al (2023) Prediction of methicillin-resistant Staphylococcus aureus and carbapenem-resistant Klebsiella pneumoniae from flagged blood cultures by combining rapid Sepsityper MALDI-TOF mass spectrometry with machine learning. *Int J Antimicrob Agents* 62:106994. <https://doi.org/10.1016/j.ijantimicag.2023.106994>
79. Abu-Aqil G, Suleiman M, Sharaha U, Riesenberger K, Lapidot I, Huleihel M et al (2023) Fast identification and susceptibility determination of E. coli isolated directly from patients' urine using infrared-spectroscopy and machine learning. *Spectrochim Acta A Mol Biomol Spectrosc* 285:121909. <https://doi.org/10.1016/j.saa.2022.121909>
80. Candela A, Arroyo MJ, Sánchez-mollada Á, Méndez G, Quiroga L, Ruiz A et al (2022) Rapid and reproducible MALDI-TOF-based method for the detection of vancomycin-resistant enterococcus faecium using classifying algorithms. *Diagnostics* 12:1–13
81. Wang J, Xia C, Wu Y, Tian X, Zhang K, Wang Z (2022) Rapid detection of carbapenem-resistant Klebsiella pneumoniae using machine learning and MALDI-TOF MS platform. *Infect Drug Resist* 15:3703–3710
82. Agbaria AH, Rosen GB, Lapidot I, Rich DH, Mordechai S, Kapelushnik J et al (2020) Rapid diagnosis of infection etiology in febrile pediatric oncology patients using infrared spectroscopy of leukocytes. *J Biophotonics* 13:1–12
83. Zhang YM, Tsao MF, Chang CY, Lin KT, Keller JJ, Lin HC (2023) Rapid identification of carbapenem-resistant Klebsiella pneumoniae based on matrix-assisted laser desorption ionization time-of-flight mass spectrometry and an artificial neural network model. *J Biomed Sci.* 30:1–10. <https://doi.org/10.1186/s12929-023-00918-2>
84. Tsai WC, Liu CF, Ma YS, Chen CJ, Lin HJ, Hsu CC et al (2023) Real-time artificial intelligence system for bacteremia prediction in adult febrile emergency department patients. *Int J Med Inform* 178:105176. <https://doi.org/10.1016/j.ijmedinf.2023.105176>
85. Ananda-Rajah MR, Bergmeir C, Petitjean F, Slavin MA, Thursky KA, Webb GI (2017) Toward electronic surveillance of invasive mold diseases in hematology-oncology patients: an expert system combining natural language processing of chest computed tomography reports, microbiology, and antifungal drug data. *JCO Clin Cancer Inform.* 1:1–10
86. Kim C, Choi YH, Choi JY, Choi HJ, Park RW, Rhie SJ (2023) Translation of machine learning-based prediction algorithms to personalised empiric antibiotic selection: a population-based cohort study. *Int J Antimicrob Agents.* 62:106966. <https://doi.org/10.1016/j.ijantimicag.2023.106966>
87. Feretzakis G, Loupelis E, Sakagianni A, Kalles D, Lada M, Christopoulos C et al (2020) Using machine learning algorithms to predict antimicrobial resistance and assist empirical treatment. *Stud Health Technol Inform.* 272:75–8
88. Feretzakis G, Sakagianni A, Loupelis E, Kalles D, Martsoukou M, Skarmoutsou N et al (2021) Using machine learning to predict antimicrobial resistance of acinetobacter baumannii, klebsiella pneumoniae and pseudomonas aeruginosa strains. *Public Health Inform Proc MIE 2021:*43–47
89. Page PS, Greenway GP, Ammanuel SG, Brooks NP (2022) Development and validation of a predictive model for failure of medical management in spinal epidural abscesses. *Neurosurgery* 91:422–426
90. Shang JS, Lin YSE, Goetz AM (2000) Diagnosis of MRSA with neural networks and logistic regression approach. *Health Care Manag Sci* 3:287–297
91. Abu-Aqil G, Suleiman M, Sharaha U, Lapidot I, Huleihel M, Salman A (2023) Instant detection of extended-spectrum β-lactamase-producing bacteria from the urine of patients using infrared spectroscopy combined with machine learning. *Analyst* 148:1130–1140
92. Feretzakis G, Loupelis E, Sakagianni A, Kalles D, Martsoukou M, Lada M et al (2020) Using machine learning techniques to aid empirical antibiotic therapy decisions in the intensive care unit of a general hospital in Greece. *Antibiotics* 9:50
93. Kanjilal S, Oberst M, Boominathan S, Zhou H, Hooper DC, Sontag D (2020) A decision algorithm to promote outpatient antimicrobial stewardship for uncomplicated urinary tract infection. *Sci Transl Med* 12:568
94. Chan E, Leroi M (2021) Evaluation of the VITEK 2 Advanced Expert System performance for predicting resistance mechanisms in Enterobacterales acquired from a hospital-based screening program. *Pathology* 53:763–767
95. Azar AT, El-Metwally SM (2013) Decision tree classifiers for automated medical diagnosis. *Neural Comput Appl* 23:2387–2403
96. Dos Santos CFG, Papa JP (2022) Avoiding overfitting: a survey on regularization methods for convolutional neural networks. *ACM Comput Surv.* 54:1–25
97. Bailly A, Blanc C, Francis É, Guillotin T, Jamal F, Wakim B et al (2022) Effects of dataset size and interactions on the prediction performance of logistic regression and deep learning models. *Comput Methods Programs Biomed.* 213:106504. <https://doi.org/10.1016/j.cmpb.2021.106504>
98. Bailey J, Oliveri A, Levin E (2013) A review of feature reduction techniques in neuroimaging. *Bone* 23:1–7

99. Ahmed SF, Alam MS Bin, Hassan M, Rozbu MR, Ishtiak T, Rafa N et al (2023) Deep learning modelling techniques: current progress, applications, advantages, and challenges. *Artif Intell Rev*. Springer Netherlands. <https://doi.org/10.1007/s10462-023-10466-8>
100. Szmigiel A, Apel DB, Pu Y, Pourrahimian Y, Dehghanpour H (2024) Exploring machine learning techniques for open stope stability prediction: a comparative study and feature importance analysis. *Rock Mech Bull*. 100146. <https://doi.org/10.1016/j.rockmb.2024.100146>
101. Zion I, Ozuomba S, Asuquo P (2020) An Overview of Neural Network Architectures for Healthcare. 2020 International Conference in Mathematics, Computer Engineering and Computer Science, ICMCECS 2020. 10
102. Pandey SK, Janghel RR (2019) Recent deep learning techniques, challenges and its applications for medical healthcare system: a review. *Neural Process Lett*. 50:1907–35. <https://doi.org/10.1007/s11063-018-09976-2>
103. Perez H, Tah JHM (2020) Improving the accuracy of convolutional neural networks by identifying and removing outlier images in datasets using t-SNE. *Mathematics*. 8:662
104. Chen RC, Dewi C, Huang SW, Caraka RE (2020) Selecting critical features for data classification based on machine learning methods. *J Big Data*. 7. <https://doi.org/10.1186/s40537-020-00327-4>
105. Munappy AR, Bosch J, Olsson HH, Arpteg A, Brinne B (2022) Data management for production quality deep learning models: challenges and solutions. *J Syst Softw*. 191:111359. <https://doi.org/10.1016/j.jss.2022.111359>
106. Tang R, Luo R, Tang S, Song H, Chen X (2022) Machine learning in predicting antimicrobial resistance: a systematic review and meta-analysis. *Int J Antimicrob Agents*. 60:106684. <https://doi.org/10.1016/j.ijantimicag.2022.106684>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.